Cellular Microparticles In The Pathogenesis Of Pulmonary Hypertension

Nicolas Amabile * ^{1,2,3}, Christophe Guignabert * ^{3,4,5}, David Montani ^{3,4,5,6}, Yerem Yeghiazarians ⁷, Chantal M. Boulanger ^{1,2}, Marc Humbert ^{3,4,5,6}

Short title: Microvesicles in pulmonary hypertension

- *: Both authors contributed equally.
- 1: INSERM U970, Paris Cardiovascular Research Center, Paris, France
- 2: Université Paris Descartes, Sorbonne Paris Cité, UMR-S970, Paris, France
- Centre Chirurgical Marie Lannelongue, Cardiology Department & Research Department,
 Le Plessis Robinson, France
- 4: INSERM U999, LabEx LERMIT, Centre Chirurgical Marie Lannelongue, Le Plessis Robinson, France
- 5: Univ Paris-Sud, Faculté de médecine, Kremlin-Bicêtre, France
- 6: Centre de Référence de l'Hypertension Pulmonaire Sévère, Service de Pneumologie et Réanimation Respiratoire, DHU Thorax Innovation, Assistance Publique Hôpitaux de Paris, Hôpital de Bicêtre, Le Kremlin-Bicetre, France.
- 7: Division of Cardiology, Department of Medicine, University of California San Francisco, San Francisco, California, United States of America.

*Address correspondence to either:

Marc Humbert, MD, PhD, INSERM U999, Centre chirurgical Marie Lannelongue, 133

Avenue de la Résistance, 92350 Le Plessis-Robinson, France. Tel: 33-1-40 94 88 33;

Fax: 33-1-40 94 25 22; e-mail: marc.humbert@abc.aphp.fr

or Chantal M. Boulanger, PhD, INSERM - U970 , 56, rue Leblanc, 75373 Paris cedex 15, France . Tel: +331.53.98.80.86 ; e-mail : chantal.boulanger@inserm.fr

Manuscript word count: 5851 words

Abstract

Pulmonary hypertension (PH) is a fatal disease with no treatment options characterized by

elevated pulmonary vascular resistance and secondary right ventricular failure. The etiology

of PAH is multiple and its pathogenesis is complex. Although the exact role of cellular

microparticles (MPs) remains partially understood, there is increasing evidence to suggest an

active role for MPs in PH pathophysiology. Patients with PH exhibited higher circulating

levels of MPs compared to controls subjects and in vitro or in vivo generated MPs can induce

endothelial dysfunction, interfere with coagulation pathways or modulate inflammatory

phenomenon. Whether or not these new conveyors of biological information contribute to the

acquisition and/or maintenance to the altered endothelial phenotype is unexplored in PH and

requires further study.

Word count: 119 words

Key words: Pulmonary arterial hypertension; Microvesicles; Endothelial dysfunction;

Pulmonary vascular remodelling; Cellular cross talk.

1

Introduction

Pulmonary hypertension (PH) is characterized by widespread obstruction and obliteration of pulmonary arterioles, leading to a progressive elevation in pulmonary vascular resistance and subsequent right-heart failure and death. The exact mechanisms of pulmonary arterial remodelling that lead to the onset and progression of PH are still largely unclear. However, many disease-predisposing factors and/or contributing factors have been identified, including inflammation, endothelial cell dysfunction, aberrant vascular wall cell proliferation and mutations in the *bone morphogenetic protein-receptor type 2 (Bmpr2)* gene [1-3].

Precise and dynamic cross talk between cells is critical for cellular behaviour, including for cell proliferation, apoptosis, differentiation, migration and survival, and thus crucial for proper tissue organization and homeostasis; abnormal cellular cross talk can lead to the development of cancer and PH [4, 5]. These well-regulated processes to transfer information between cells occur through direct cell-cell contact, various soluble bioactive factors and through cellular microparticles (MPs) [6, 7].

MPs are plasma membrane vesicle fragments (between 0.1 and 1 µm in size) released from various cell types during activation by agonists or physical or chemical stress, including apoptosis [6, 7]. MPs in body fluids constitute a heterogeneous population, differing in cellular origin (endothelial cells (ECs), platelets, leukocytes and erythrocytes), number, size, antigenic composition and functional properties. Their concentration increases specifically and differently during various pathological states, including cardiovascular diseases (Table 1) [8]. Indeed, patients with PH (Group 1 and 3 with no associated cardiovascular disease) exhibit higher circulating levels of MPs compared to control subjects [9-11]. This increase might indicate a potential involvement of MPs in the disease onset and its progression. This article reviews the evidence supporting potential roles and importance for MPs in the pathogenesis of PH.

General considerations on Microparticles

Detectable levels of MPs of different cellular origin (platelets, red blood cells, leukocytes, endothelial cells) circulate in the plasma of healthy subjects (Figure 1B). Circulating MPs levels increase in a wide range of cardiovascular diseases, including uncontrolled cardiovascular risk factors, stable and unstable atherosclerotic lesions, heart failure, arrhythmias and inflammatory vascular diseases (Table 1) [8]. The presence of MPs in plasma reflects an active balance between MP generation and clearance. Both phagocytic activities present in the liver and the spleen contribute to the removal of MPs from plasma of healthy animals [12, 13]. Changes in circulating levels of MPs might bear important clinical information in healthy subjects and in patients with cardiovascular disorders. MPs display various bioactive substances and receptors on their surface and harbour a concentrated set of cytokines, signaling proteins, lipids, and nucleic acids. The composition of MPs, which subtends their biological effect, is a function of the parent cell they stem from as well as the stimulus used for their generation. Thus, the amount and nature of circulating MPs influence their beneficial or deleterious biological effects in physiological or pathological conditions. Indeed, various experimental evidences reported significant interrelations between circulating MPs (either generated in vivo or in vitro) and other protagonists of disrupted vascular homeostasis. MPs can modulate vascular reactivity (particularly endothelial dependent relaxation), interfere with coagulation pathways, enhance or decrease inflammatory phenomenon and influence neo-angiogenesis process. A large part of our knowledge regarding MPs is based on data issued from the research on athero-thrombotic disease [14]. However, recent data shed the light on their role as vectors of cellular cross talk with implications in the pathogenesis of non-atherosclerotic cardiovascular conditions, including pulmonary vascular diseases (Table 1).

Circulating Microparticles as Regulators of Endothelial Dysfunction

Endothelial dysfunction is a pivotal element in the development and progression of PH, irrespective of disease origin [15]. The dysfunctional endothelium displays to varying degrees an imbalanced production of several mediators (vasoconstrictors versus vasodilators; activators versus inhibitors of smooth muscle cell growth; proinflammatory versus antiinflammatory signals; prothrombotic versus antithrombotic substances) leading towards an excess of vasoconstriction, smooth muscle hyperplasia and pulmonary vascular remodeling [1-3]. In addition, disturbances of the normal balance between endothelial cell proliferation and endothelial cell apoptosis have been reported in idiopathic PAH [16, 17]. Indeed, accumulating evidence supports the concept that increased endothelial apoptosis at early stage and decreased endothelial apoptosis at later stages of the disease could contribute to PAH [18]. Consistent with this hypothesis, we have recently reported that primary pulmonary endothelial cells generated from PAH lung specimens exhibit various intrinsic abnormalities and present a modified pro-proliferative, apoptotic-resistant phenotype [4, 17, 19]. Although we have shown that increased activity of the FGF2 autocrine loop is among the mechanisms needed to acquire this altered endothelial phenotype in PAH [17], the exact nature of pulmonary endothelial cell modification during PAH and the balance between apoptotic and anti-apoptotic phenotypes remain only partially understood. In addition, alterations of endothelial cell monolayer integrity in mice, such as a selective disruption of the endothelial peroxisome proliferator-activated receptor (PPAR)y or of the endothelial BMPR2 signaling, are sufficient to predispose to PH [20, 21].

Different groups have reported increased concentrations of different subsets of circulating endothelial MPs (EMPs) in PH patients compared to controls subjects [9-11], supporting the presence of an increased structural damage with enhanced cellular activation and/or apoptosis. Indeed, the levels of PECAM+ and VE-cadherin+ EMPs were also correlated with several hemodynamics pulmonary vascular remodeling parameters [10] (Figure 1) and inflammation biomarkers [11]. The etiology of the disease appears also to

influence the MPs levels: Amabile et al. reported that PAH patients had higher levels of EMPs compared to chronic pulmonary disease related PH subjects [10], whereas Diehl et al. observed higher levels of EMPs bearing E-selectin in thrombo-embolic PH compared to non-thrombo-embolic PH subjects [11].

The origin of these circulating MPs (systemic vs. pulmonary endothelium) is still a matter of debate. Although a trans-pulmonary gradient of CD105+ (endoglin) EMPs has been observed, suggesting a local release of MPs within the pulmonary vascular bed, no significant change in CD31+ (PECAM), CD144+ (VE-Cadherin) and CD62e+ (E-selectin) EMP levels was observed through the pulmonary vasculature [9, 10]. These findings, observed in small samples of patients with various PH origins, could reflect the heterogeneity of the endothelial changes among patients with pulmonary vascular disease, depending on the underlying etiology or the lesion development stage (early vs. late stage). These findings might support the notion that some specific EMP subpopulations could be released by either systemic or pulmonary endothelium. Finally, it cannot be excluded that a lack of sensitivity of the measurement methods for detection of mild variations in concentrations of MPs could also explain these results.

The circulating MPs are essential vectors of biological informations and are involved in cellular communication through a paracrine or endocrine action [22]. Cellular effects of MPs are directly influenced by the various antigens harbored at their surface, which is a function of the cellular origin and the stimulus involved in the vesiculation process [8]. *In vitro* or *in vivo* generated MPs can induce endothelial dysfunction, interfere with coagulation pathways or modulate inflammatory phenomenon [8]. Accordingly, Tual-Chalot *et al.* reported that circulating MPs isolated from hypoxia-induced PH rats could affect the vascular tone [23]. They showed that these MPs, mainly of platelet and erythrocyte origin, impaired the endothelium-dependent vaso-relaxation in pulmonary arteries *ex vivo* and decreased the nitric oxide (NO) production by pulmonary endothelial cells through increase in oxidative stress [23]. The authors observed comparable results in aorta and pulmonary arteries [23].

Taken together, all these experimental findings support the notion that circulating MPs could contribute to both initiation and/or amplification of the pulmonary endothelial dysfunction during PH and encourage further studies of the precise pathogenic mechanism underlying this phenomenon. Moreover, we cannot exclude that the generation of MPs and their migration to the systemic circulation could mediate a cross-talk between pulmonary and systemic endothelium and therefore explain the peripheral endothelial dysfunction observed during PH [21, 24, 25].

Finally, measurement of circulating EMPs might also represent a potential new prognostic tool in patients with PH. Preliminary data reported that baseline values of circulating CD62e+ MPs, measured before initiation of any specific therapy, predicted occurrence of adverse outcome (death+ readmission for right heart failure) in a small cohort of patients with mixed etiologies of PH [26]. Whereas these results were not observed for other EMP subsets (CD144+ and CD31+ EMPs) and should be confirmed in a larger population sample, they are in line with data reported in coronary artery disease [27], heart failure [28] or chronic end-stage renal failure [29] and thus confirm their potential role as a biomarker for outcome prediction in patients with cardiovascular diseases [30].

Circulating Microparticles as Regulators of Inflammation

It is now well established that inflammation plays an important role in PH in humans: histological studies of the PAH lung specimens demonstrate a prominent perivascular infiltration composed of macrophages, dendritic cells, T and B lymphocytes, and mast cells; circulating levels of certain cytokines and chemokines are elevated including interleukin (IL)-1β, IL-6, IL-8, CX3CL1, CCL2, CCL5 and tumor necrosis factor (TNF)-α serum levels, and these may correlate with a worse clinical outcome. Evidence supporting a role for inflammation in the progression of the disease have been reviewed recently by Price *et al.* [31]. However, the initial process triggering inflammation onset remains unknown.

Cellular MPs are key actors in the inflammatory process because they contribute to the endothelial production of various pro-inflammatory cytokines and chemokines. MPs have been shown to stimulate the release of IL-1ß, IL-6, CCL2, and induce the expression of intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1) and E-selectin [32-35]. In addition, MPs provide a preferential substrate for non-pancreatic secretory phospholipase A2 (sPLA2) and thus a source of lysophosphatic acid, a potent pro-inflammatory mediator and platelet agonist [36]. Furthermore, a recent study has shown that MPs from septic shock patients increase the expression of endothelial and inducible nitric oxide synthase (iNOS), cyclooxygenase-2, and nuclear factor-kB in heart and lung [37]. In addition, Densmore *et al.* have demonstrated that endothelium-derived MPs sequestration in the lung is capable of initiating inflammatory cascades at pathophysiologically relevant concentrations [38, 39]. Altogether, these findings support the notion that circulating MPs could actively contribute to the initiation and/or amplification of the pulmonary vascular inflammation in the pathogenesis of PH.

Procoagulant Properties of Microparticles

Thrombosis is a key mechanism in onset and progression of cardiovascular diseases, including venous thromboembolic disease, atherosclerosis and PH. Chronic thromboembolic PH results from the obstruction of the pulmonary arteries by clot fragments migrated from deep venous thrombosis [40]. On the other hand, PAH pathological specimens often display thrombotic lesions in the absence of clinical or pathological evidence of pulmonary embolism [41], suggesting an *in situ* clotting phenomenon [42]. Furthermore, PH is associated with a hypercoagulable phenotype [43] that includes vascular up-regulation of tissue factor [44] and an increase in circulating levels of Von Willebrand factor or fibrinopeptide A [45].

MPs are involved in thrombus generation by different mechanisms. MPs can provide the anionic phospholipid surface necessary for the assembly of clotting enzyme complexes (VIIIa and Xa) promoting thrombin generation. In some cases, MPs also directly expose

tissue factor, the main cellular initiator of blood coagulation, at their surface. Furthermore, MPs can also harbour other functional membrane or cytoplasmic effectors (such as selectins, GPIIbIIIa, GPIb, von Willebrand factor, arachidonic acid, thromboxane A2, etc.) that can promote prothrombotic responses [22]. Therefore, MPs can promote thrombin generation *in vitro*, but also favour thrombus propagation *in vivo* [46] or transfer their procoagulant potential to target cells. The thrombogenic potential of MPs has been demonstrated for several subpopulations, including platelets [47], endothelial [48, 49] or leukocytes derived MPs [50].

Most of our knowledge regarding MPs' procoagulant activity derives from data observed in atherosclerosis, cardiac surgery and conditions associated with cardiomyocyte inflammatory stimulation, such as cardiac ischemia.

An increase in circulating global procoagulants as well as TF+ MPs (MPs bearing tissue factor) levels has been reported in patients with PAH [9]. Furthermore, the levels of TF+ MPs and von Willebrand factors were correlated. Interestingly, the highest levels of TF+- MPs were observed in patients with functional class III-IV and/or the weakest 6-min test performance, suggesting a potential correlation between MPs and functional status. Comparable results were observed in patients with recurrent deep venous thrombosis, a potential trigger for acute pulmonary embolism development [51].

The precise molecular mechanisms governing these biological effects of MPs are still under investigation, but several elements support the notion that procoagulant MPs might interfere with the coagulation process in PH patients and promote the disease development in different ways. High levels of procoagulant circulating MPs could favour recurrent deep venous thrombosis [52], which will in turn create silent pulmonary embolism and ultimately lead to post-thromboembolic PH [40, 53]. On the other hand, the plexiform lesions and the dysfunctional endothelium could release procoagulant MPs, which could lead to propagation of the thrombotic process among the pulmonary vessels. The resulting local thrombosis could worsen the microvascular obstruction and thus increase pulmonary artery pressure

(PAP). This hypothesis is supported by the results from Bakouboula *et al.*, who observed an increase in the concentration of the global number of procoagulant MPs between the jugular vein and the wedged pulmonary artery, suggesting a local release of procoagulant microvesicles [9]. However, the authors reported no significant transpulmonary gradient of TF+ MPs, which might be explained by the small size of the group (n=20) and the heterogeous etiologies of PH [9]. Moreover, proteomics analysis in experimental models of post thromboembolic PH have shown that this condition was associated with an increased expression of coagulation proteins carried by circulating MPs [54].

Potential implications of MPs in PH pathophysiology and remaining questions

In the light of the above-mentioned data, the involvement of MPs in PH pathogenesis seems to be, at least, plausible. Experimental and clinical data suggest multiple roles of MPs in the progression of the disease and one might hypothesize a possible global scheme (Figure 1C). Pro-coagulant MPs can enhance local and systemic thrombogenicity leading to pulmonary artery clot. Moreover, pro-inflammatory MPs could also increase endothelial inflammation, local cytokine and growth factors release. Furthermore, MPs might also favor endothelial cell proliferation. Finally, circulating MPs can enhance endothelial dysfunction, decrease NO release and deteriorate the vasodilation/vasoconstriction humoral balance. Altogether, these effects can support pulmonary vessel remodeling and vascular obstruction and should encourage further *in vivo* and *in vitro* studies to precise their pathogenic mechanisms. In addition, a very recent study has reported that epoprostenol could inhibit human platelet-leukocyte mixed conjugate and platelet microparticle formation in whole blood, demonstrating that treatments might interfere with MPs and some of their biological role [55]. Since our knowledge on microparticles properties *in vivo* might also be influenced by the current methods used for their isolation and their measurements, there is a need to

establish a consensus regarding the standard centrifugation method to prepare poor platelets plasma from whole blood samples in order to determine MPs phenotypes and levels. Nowadays, the appropriate method for MPs measurement is still a matter of debate [56]. Flow cytometry methods have been widely used during the past 20 years, need small samples volumes and allow complete phenotypical analysis. However, flow cytometry doesn't analyze MPs function and is limited by its poor sensitivity, resulting from the biophysical and optical properties of laser interaction with small vesicles (<500 µm). The "capture" and other purely biological methods inform on MPs function but don't provide any direct quantification. Dynamic light scattering, atomic force microscopy, automated flow through single-particle optical microscopy or impedance based flow cytometry represent promising methods for the future, allowing more accurate MPs quantification, but these methods are not currently adapted for clinical research as they are limited by their cost or their availability.

In addition to these potential pathogenic roles of MPs in PH, MPs are well-known modulators of different stages of angiogenesis. These particular effects of MPs depend on the stimulation and activation status of the cell from which they originate from and also on their concentration [57, 58]. Indeed, MPs from platelets display pro-angiogenic properties by promoting capillary-like structures and pro-angiogenic factor production [59]. By contrast, lymphocyte- and endothelial-derived MPs exhibit pro- or anti-angiogenic properties depending on the stimuli used for their production [57, 60-63]. These additional properties of MPs reinforce the idea that they could be potential key actors in PH pathogenesis in the early-, middle- and/or late-stage of the disease.

Whether or not these new conveyors of biological information contribute to the acquisition and/or maintenance to the altered endothelial phenotype is not fully explored in PH and requires further study. Thus, several questions remain regarding their nature and true importance in this disease. What are the compositions of MPs in PH and what are their functions? What causes increased amounts of circulating MPs in PH? Are they beneficial or

deleterious in PH? Are they useful for disease recurrence, prognosis, and monitoring of therapeutic efficacy? By which potential mechanism(s) do MPs initiate and propagate PH?

Conclusion

Patients with PH exhibit higher circulating levels of MPs compared to control subjects and it is unclear whether these new conveyors of biological information contribute to the acquisition and/or maintenance to the altered endothelial phenotype or are just a consequence of the disease progression. However, there is strong accumulating evidence that MPs are key protagonists of the disease and they could serve as a worthy biomarker for prognosis stratification and therapeutic efficiency monitoring. Therefore, a better understanding of their importance in the disease is essential and efforts should be made in the future to improve our knowledge by further exploring cellular aspects as well as clinical implications of MPs in pulmonary hypertension.

Conflict of interest: The authors declare that no conflict of interest exists.

Disclosures: None of the authors have any disclosures related to conflict of interest.

References

- Humbert M, Morrell NW, Archer SL, Stenmark KR, MacLean MR, Lang IM, Christman BW, Weir EK, Eickelberg O, Voelkel NF, Rabinovitch M. Cellular and molecular pathobiology of pulmonary arterial hypertension. *J Am Coll Cardiol* 2004; 43: 13S-24S.
- Morrell NW, Adnot S, Archer SL, Dupuis J, Jones PL, MacLean MR, McMurtry IF, Stenmark KR, Thistlethwaite PA, Weissmann N, Yuan JX, Weir EK. Cellular and molecular basis of pulmonary arterial hypertension. *J Am Coll Cardiol* 2009; 54: S20-31.
- 3. Rabinovitch M. Molecular pathogenesis of pulmonary arterial hypertension. *The Journal of clinical investigation* 2008; 118: 2372-2379.
- 4. Eddahibi S, Guignabert C, Barlier-Mur AM, Dewachter L, Fadel E, Dartevelle P, Humbert M, Simonneau G, Hanoun N, Saurini F, Hamon M, Adnot S. Cross talk between endothelial and smooth muscle cells in pulmonary hypertension: critical role for serotonin-induced smooth muscle hyperplasia. *Circulation* 2006; 113: 1857-1864.
- 5. Xu R, Mao JH. Gene transcriptional networks integrate microenvironmental signals in human breast cancer. *Integr Biol (Camb)* 2011; 3: 368-374.
- 6. Diamant M, Tushuizen ME, Sturk A, Nieuwland R. Cellular microparticles: new players in the field of vascular disease? *European journal of clinical investigation* 2004; 34: 392-401.
- 7. Freyssinet JM. Cellular microparticles: what are they bad or good for? *Journal of thrombosis and haemostasis : JTH* 2003; 1: 1655-1662.
- 8. Amabile N, Rautou PE, Tedgui A, Boulanger CM. Microparticles: key protagonists in cardiovascular disorders. *Semin Thromb Hemost* 2010; 36: 907-916.
- Bakouboula B, Morel O, Faure A, Zobairi F, Jesel L, Trinh A, Zupan M, Canuet M,
 Grunebaum L, Brunette A, Desprez D, Chabot F, Weitzenblum E, Freyssinet JM,
 Chaouat A, Toti F. Procoagulant membrane microparticles correlate with the severity of

- pulmonary arterial hypertension. *American journal of respiratory and critical care medicine* 2008; 177: 536-543.
- Amabile N, Heiss C, Real WM, Minasi P, McGlothlin D, Rame EJ, Grossman W, De Marco T, Yeghiazarians Y. Circulating endothelial microparticle levels predict hemodynamic severity of pulmonary hypertension. *American journal of respiratory and critical care medicine* 2008; 177: 1268-1275.
- 11. Diehl P, Aleker M, Helbing T, Sossong V, Germann M, Sorichter S, Bode C, Moser M. Increased platelet, leukocyte and endothelial microparticles predict enhanced coagulation and vascular inflammation in pulmonary hypertension. *Journal of Thrombosis and Thrombolysis* 2011; 31: 173-179.
- Dasgupta SK, Abdel-Monem H, Niravath P, Le A, Bellera RV, Langlois K, Nagata S,
 Rumbaut RE, Thiagarajan P. Lactadherin and clearance of platelet-derived microvesicles. *Blood* 2009; 113: 1332-1339.
- 13. Willekens FL, Werre JM, Kruijt JK, Roerdinkholder-Stoelwinder B, Groenen-Dopp YA, van den Bos AG, Bosman GJ, van Berkel TJ. Liver Kupffer cells rapidly remove red blood cell-derived vesicles from the circulation by scavenger receptors. *Blood* 2005; 105: 2141-2145.
- Rautou P-E, Vion A-Cm, Amabile N, Chironi G, Simon A, Tedgui A, Boulanger CM.
 Microparticles, Vascular Function, and Atherothrombosis. *Circulation Research* 2011;
 109: 593-606.
- 15. Budhiraja R, Tuder RM, Hassoun PM. Endothelial dysfunction in pulmonary hypertension. *Circulation* 2004; 109: 159-165.
- 16. Masri FA, Xu W, Comhair SA, Asosingh K, Koo M, Vasanji A, Drazba J, Anand-Apte B, Erzurum SC. Hyperproliferative apoptosis-resistant endothelial cells in idiopathic pulmonary arterial hypertension. *American journal of physiology Lung cellular and molecular physiology* 2007; 293: L548-554.
- Tu L, Dewachter L, Gore B, Fadel E, Dartevelle P, Simonneau G, Humbert M, Eddahibi
 S, Guignabert C. Autocrine fibroblast growth factor-2 signaling contributes to altered

- endothelial phenotype in pulmonary hypertension. *American journal of respiratory cell* and molecular biology 2011; 45: 311-322.
- 18. Sakao S, Taraseviciene-Stewart L, Lee JD, Wood K, Cool CD, Voelkel NF. Initial apoptosis is followed by increased proliferation of apoptosis-resistant endothelial cells.

 FASEB journal: official publication of the Federation of American Societies for Experimental Biology 2005; 19: 1178-1180.
- 19. Tu L, De Man FS, Girerd B, Huertas A, Chaumais MC, Lecerf F, Francois C, Perros F, Dorfmuller P, Fadel E, Montani D, Eddahibi S, Humbert M, Guignabert C. A Critical Role for p130Cas in the Progression of Pulmonary Hypertension in Humans and Rodents. American journal of respiratory and critical care medicine 2012.
- 20. Guignabert C, Alvira CM, Alastalo TP, Sawada H, Hansmann G, Zhao M, Wang L, El-Bizri N, Rabinovitch M. Tie2-mediated loss of peroxisome proliferator-activated receptor-gamma in mice causes PDGF receptor-beta-dependent pulmonary arterial muscularization. *American journal of physiology Lung cellular and molecular physiology* 2009; 297: L1082-1090.
- 21. Hong KH, Lee YJ, Lee E, Park SO, Han C, Beppu H, Li E, Raizada MK, Bloch KD, Oh SP. Genetic ablation of the BMPR2 gene in pulmonary endothelium is sufficient to predispose to pulmonary arterial hypertension. *Circulation* 2008; 118: 722-730.
- Morel O, Toti F, Hugel B, Bakouboula B, Camoin-Jau L, Dignat-George F, Freyssinet J-M. Procoagulant Microparticles. Disrupting the Vascular Homeostasis Equation?
 Arterioscler Thromb Vasc Biol 2006; 26: 2594-2604.
- 23. Tual-Chalot S, Guibert C, Muller B, Savineau J-P, Andriantsitohaina R, Martinez MC. Circulating Microparticles from Pulmonary Hypertensive Rats Induce Endothelial Dysfunction. *Am J Respir Crit Care Med* 2010; 182: 261-268.
- 24. Friedman D, Szmuszkovicz J, Rabai M, Detterich JA, Menteer J, Wood JC. Systemic endothelial dysfunction in children with idiopathic pulmonary arterial hypertension correlates with disease severity. *The Journal of heart and lung transplantation : the*

- official publication of the International Society for Heart Transplantation 2012; 31: 642-647.
- 25. Peled N, Bendayan D, Shitrit D, Fox B, Yehoshua L, Kramer MR. Peripheral endothelial dysfunction in patients with pulmonary arterial hypertension. *Respiratory medicine* 2008; 102: 1791-1796.
- 26. Amabile N, Heiss C, Chang V, Angeli FS, Damon L, Rame EJ, McGlothlin D, Grossman W, De Marco T, Yeghiazarians Y. Increased CD62e(+) endothelial microparticle levels predict poor outcome in pulmonary hypertension patients. *J Heart Lung Transplant* 2009; 28: 1081-1086.
- 27. Nozaki T, Sugiyama S, Koga H, Sugamura K, Ohba K, Matsuzawa Y, Sumida H, Matsui K, Jinnouchi H, Ogawa H. Significance of a multiple biomarkers strategy including endothelial dysfunction to improve risk stratification for cardiovascular events in patients at high risk for coronary heart disease. *J Am Coll Cardiol* 2009; 54: 601-608.
- 28. Nozaki T, Sugiyama S, Sugamura K, Ohba K, Matsuzawa Y, Konishi M, Matsubara J, Akiyama E, Sumida H, Matsui K, Jinnouchi H, Ogawa H. Prognostic value of endothelial microparticles in patients with heart failure. *European Journal of Heart Failure* 2010; 12: 1223-1228.
- 29. Amabile N, Guerin AP, Tedgui A, Boulanger CM, London GM. Predictive value of circulating endothelial microparticles for cardiovascular mortality in end-stage renal failure: a pilot study. *Nephrology, dialysis, transplantation : official publication of the European Dialysis and Transplant Association European Renal Association* 2011.
- 30. Amabile N, Boulanger CM. Circulating microparticle levels in patients with coronary artery disease: a new indicator of vulnerability? *European Heart journal* 2011; 32: 1958-1960.
- Price LC, Wort SJ, Perros F, Dorfmuller P, Huertas A, Montani D, Cohen-Kaminsky S,
 Humbert M. Inflammation in pulmonary arterial hypertension. *Chest* 2012; 141: 210-221.

- 32. MacKenzie A, Wilson HL, Kiss-Toth E, Dower SK, North RA, Surprenant A. Rapid secretion of interleukin-1beta by microvesicle shedding. *Immunity* 2001; 15: 825-835.
- 33. Nomura S, Tandon NN, Nakamura T, Cone J, Fukuhara S, Kambayashi J. High-shear-stress-induced activation of platelets and microparticles enhances expression of cell adhesion molecules in THP-1 and endothelial cells. *Atherosclerosis* 2001; 158: 277-287.
- 34. Rautou PE, Leroyer AS, Ramkhelawon B, Devue C, Duflaut D, Vion AC, Nalbone G, Castier Y, Leseche G, Lehoux S, Tedgui A, Boulanger CM. Microparticles from human atherosclerotic plaques promote endothelial ICAM-1-dependent monocyte adhesion and transendothelial migration. *Circulation research* 2011; 108: 335-343.
- 35. Vince RV, McNaughton LR, Taylor L, Midgley AW, Laden G, Madden LA. Release of VCAM-1 associated endothelial microparticles following simulated SCUBA dives. *Eur J Appl Physiol* 2009; 105: 507-513.
- 36. Fourcade O, Simon MF, Viode C, Rugani N, Leballe F, Ragab A, Fournie B, Sarda L, Chap H. Secretory phospholipase A2 generates the novel lipid mediator lysophosphatidic acid in membrane microvesicles shed from activated cells. *Cell* 1995; 80: 919-927.
- 37. Mastronardi ML, Mostefai HA, Meziani F, Martinez MC, Asfar P, Andriantsitohaina R. Circulating microparticles from septic shock patients exert differential tissue expression of enzymes related to inflammation and oxidative stress. *Crit Care Med* 2011; 39: 1739-1748.
- 38. Buesing KL, Densmore JC, Kaul S, Pritchard KA, Jr., Jarzembowski JA, Gourlay DM, Oldham KT. Endothelial microparticles induce inflammation in acute lung injury. *The Journal of surgical research* 2011; 166: 32-39.
- 39. Densmore JC, Signorino PR, Ou J, Hatoum OA, Rowe JJ, Shi Y, Kaul S, Jones DW, Sabina RE, Pritchard KA, Jr., Guice KS, Oldham KT. Endothelium-derived microparticles induce endothelial dysfunction and acute lung injury. *Shock* 2006; 26: 464-471.

- 40. Pengo V, Lensing AWA, Prins MH, Marchiori A, Davidson BL, Tiozzo F, Albanese P, Biasiolo A, Pegoraro C, Iliceto S, Prandoni P. Incidence of Chronic Thromboembolic Pulmonary Hypertension after Pulmonary Embolism. New England Journal of Medicine 2004; 350: 2257-2264.
- 41. Pietra GG, Edwards WD, Kay JM, Rich S, Kernis J, Schloo B, Ayres SM, Bergofsky EH, Brundage BH, Detre KM, et al. Histopathology of primary pulmonary hypertension. A qualitative and quantitative study of pulmonary blood vessels from 58 patients in the National Heart, Lung, and Blood Institute, Primary Pulmonary Hypertension Registry. Circulation 1989; 80: 1198-1206.
- 42. Bjornsson J, Edwards WD. Primary pulmonary hypertension: a histopathologic study of 80 cases. *Mayo Clinic proceedings Mayo Clinic* 1985; 60: 16-25.
- 43. Schermuly RT, Ghofrani HA, Wilkins MR, Grimminger F. Mechanisms of disease: pulmonary arterial hypertension. *Nat Rev Cardiol* 2011; 8: 443-455.
- 44. White RJ, Meoli DF, Swarthout RF, Kallop DY, Galaria II, Harvey JL, Miller CM, Blaxall BC, Hall CM, Pierce RA, Cool CD, Taubman MB. Plexiform-like lesions and increased tissue factor expression in a rat model of severe pulmonary arterial hypertension.
 American Journal of Physiology Lung Cellular and Molecular Physiology 2007; 293: L583-L590.
- 45. Johnson SR, Granton JT, Mehta S. Thrombotic Arteriopathy and Anticoagulation in Pulmonary Hypertension*. *Chest* 2006; 130: 545-552.
- 46. Furie B, Furie BC. Mechanisms of Thrombus Formation. *The New England journal of medicine* 2008; 359: 938-949.
- 47. Muller I, Klocke A, Alex M, Kotzsch M, Luther T, Morgenstern E, Zieseniss S, Zahler S, Preissner K, Engelmann B. Intravascular tissue factor initiates coagulation via circulating microvesicles and platelets. *The FASEB journal : official publication of the Federation of American Societies for Experimental Biology* 2003; 17: 476-478.

- 48. Abid Hussein MN, Boing AN, Biro E, Hoek FJ, Vogel GM, Meuleman DG, Sturk A, Nieuwland R. Phospholipid composition of in vitro endothelial microparticles and their in vivo thrombogenic properties. *Thrombosis research* 2008; 121: 865-871.
- 49. Combes V, Simon AC, Grau GE, Arnoux D, Camoin L, Sabatier F, Mutin M, Sanmarco M, Sampol J, Dignat-George F. In vitro generation of endothelial microparticles and possible prothrombotic activity in patients with lupus anticoagulant. *The Journal of clinical investigation* 1999; 104: 93-102.
- 50. Satta N, Toti F, Feugeas O, Bohbot A, Dachary-Prigent J, Eschwege V, Hedman H, Freyssinet JM. Monocyte vesiculation is a possible mechanism for dissemination of membrane-associated procoagulant activities and adhesion molecules after stimulation by lipopolysaccharide. *J Immunol* 1994; 153: 3245-3255.
- 51. Ye R, Ye C, Huang Y, Liu L, Wang S. Circulating tissue factor positive microparticles in patients with acute recurrent deep venous thrombosis. *Thrombosis research* 2011.
- 52. Bucciarelli P, Martinelli I, Artoni A, Passamonti SM, Previtali E, Merati G, Tripodi A, Mannucci PM. Circulating microparticles and risk of venous thromboembolism.

 Thrombosis research.
- 53. Kearon C. Natural History of Venous Thromboembolism. *Circulation* 2003; 107: I-22-I-30.
- 54. Watts JA, Lee YY, Gellar MA, Fulkerson MB, Hwang SI, Kline JA. Proteomics of microparticles after experimental pulmonary embolism. *Thrombosis research* 2011.
- 55. Tamburrelli C, Crescente M, Izzi B, Barisciano M, Donati MB, de Gaetano G, Cerletti C. Epoprostenol inhibits human platelet-leukocyte mixed conjugate and platelet microparticle formation in whole blood. *Thrombosis research* 2011; 128: 446-451.
- 56. Freyssinet JM, Toti F. Membrane microparticle determination: at least seeing what's being sized! *Journal of thrombosis and haemostasis : JTH* 2010; 8: 311-314.
- 57. Yang C, Mwaikambo BR, Zhu T, Gagnon C, Lafleur J, Seshadri S, Lachapelle P, Lavoie J-C, Chemtob S, Hardy P. Lymphocytic microparticles inhibit angiogenesis by

- stimulating oxidative stress and negatively regulating VEGF-induced pathways. *Am J Physiol Regul Integr Comp Physiol* 2008; 294: R467-476.
- 58. Lacroix R, Sabatier F, Mialhe A, Basire A, Pannell R, Borghi H, Robert S, Lamy E, Plawinski L, Camoin-Jau L, Gurewich V, Angles-Cano E, Dignat-George F. Activation of plasminogen into plasmin at the surface of endothelial microparticles: a mechanism that modulates angiogenic properties of endothelial progenitor cells in vitro. *Blood* 2007; 110: 2432-2439.
- 59. Brill A, Dashevsky O, Rivo J, Gozal Y, Varon D. Platelet-derived microparticles induce angiogenesis and stimulate post-ischemic revascularization. *Cardiovasc Res* 2005; 67: 30-38.
- 60. Mezentsev A, Merks RM, O'Riordan E, Chen J, Mendelev N, Goligorsky MS, Brodsky SV. Endothelial microparticles affect angiogenesis in vitro: role of oxidative stress. *Am J Physiol Heart Circ Physiol* 2005; 289: H1106-1114.
- Leroyer AS, Ebrahimian TG, Cochain C, Recalde A, Blanc-Brude O, Mees B, Vilar J, Tedgui A, Levy BI, Chimini G, Boulanger CM, Silvestre JS. Microparticles from ischemic muscle promotes postnatal vasculogenesis. *Circulation* 2009; 119: 2808-2817.
- 62. Ou Z-J, Chang F-J, Luo D, Liao X-L, Wang Z-P, Zhang X, Xu Y-Q, Ou J-S. Endothelium-derived microparticles inhibit angiogenesis in the heart and enhance the inhibitory effects of hypercholesterolemia on angiogenesis. *American Journal of Physiology Endocrinology And Metabolism* 2011; 300: E661-E668.
- 63. Reich N, Beyer C, Gelse K, Akhmetshina A, Dees C, Zwerina J, Schett G, Distler O, Distler JHW. Microparticles stimulate angiogenesis by inducing ELR+ CXC-chemokines in synovial fibroblasts. *Journal of Cellular and Molecular Medicine* 2011; 15: 756-762.

Table 1: Microparticle (MPs) involvement in cardiovascular diseases. Adapted from Amabile et al. [8]. EMPs indicates endothelial microparticles; PMPs: Platelet microparticles; LMPs: leukocyte-derived microparticles; RBCMPs: red blood cell-derived microparticles; TF: Tissue factor; PS: Phosphatidylserine.

CARDIOVASCULAR RISK FACTORS

Diabetes TF-MPs, CD8+MPs, CD66+MPs, PMPs

CD51+ EMPs, PMPs, PS+MPs

Severe systemic hypertension CD31+ EMPs, PMPs

Metabolic syndrome CD31+ EMPs, CD51+ EMPs

Hypertriglyceridemia CD31+ EMPs

ENDOTHELIAL DYSFUNCTION (ED)

Acute endothelial dysfunction CD31+EMPs, CD144+ EMPs, CD62e+ EMPs

Chronic renal failure with ED CD31+EMPs, CD144+EMPs, PMPs,

RBCMPs, PS+ MPs CD31+/PS+ MPs CD144+ EMPs CD31+/42- EMPs

Coronary artery disease with ED Diabetes with ED

Obesity with ED

ATHEROSCLEROSIS

Subclinical atherosclerosis CD11a+ LMPs

Coronary calcification CD62e+ EMPs, PMPs

Acute coronary syndrome PS+ MPs, CD31+ MPs, CD146+ EMPs CD31+ EMPs, CD51+ EMPs, PMPs

PS+ MPs, CD31+EMPs, PMPs

Acute coronary syndrome (cont.) Stable coronary artery disease CD31+ EMPs, CD51+ EMPs

Acute stroke CD105+/144+ EMPs, CD105+/54+ EMPs,

CD105+/PS+ EMPs

Cerebrovascular atherosclerosis CD62e+EMPs, CD31+ EMPs

PMPs Peripheral artery disease

HEART FAILURE & VALVULAR DISEASE

Congestive heart failure CD62e+ EMPs, CD31+ EMPs

Cyanotic congenital heart disease **PMPs**

Heart transplant rejection CD62e+ EMPs, TF+-MPs, FAS-MPs

Pulmonary hypertension CD31+ EMPs, CD144+EMPs, CD62e+EMPs,

CD45+ LMPs

CD105+ EMPs, TF+-MPs

Severe Aortic Stenosis CD62e+EMPs, CD11+ LMPs

ATRIAL FIBRILLATION PS+ MPs, PMPs

VENOUS THROMBOEMBOLISM CD31+EMPs, CD62e+ EMPs

OTHER VASCULAR DISEASES

CD105+ EMPs, CD62e+ EMPs, PMPs Inflammatory vasculitis

PS+ MPs, PMPs

Antiphospholipid syndrome CD31+/ CD51+ EMPs,

Preeclampsia CD4+MPs, CD8+MPs, CD66+ LMPs

CD31+ EMPs

Figure 1: Relationships between endothelial MPs (EMPs) and pulmonary vascular resistance in patients with pulmonary hypertension (PH) (A; adapted from[10]), the most important known types of circulating MPs, and proposed model for the role of microparticles in the PH pathogenesis (C): high circulating levels of EMPs in patients with PH (Group 1 and 3 with no associated cardiovascular disease) could: 1) contribute, at least in part, to the initiation and/ or perpetuation of the endothelial dysfunction and decreased the nitric oxide (NO) production by pulmonary endothelial cells through increase in oxidative stress; 2) interfere with coagulation pathways; and/ or 3) modulate inflammation through the stimulation of the release of various key cytokines, and/ or via induction of intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1) and E-selectin at the endothelial cell surface. EMPs indicates endothelial microparticles; IL, interleukin; CCL2, chemokine ligand 2; ICAM-1, Intercellular Adhesion Molecule-1; VCAM-1, vascular cell adhesion molecule 1; eNOS, endothelial NOS.

