

TREATMENT OF TUBERCULOSIS

UPDATE 2010

Wing Wai Yew

Tuberculosis & Chest Unit, Grantham Hospital, Hong Kong, CHINA

Christoph Lange

Division of Clinical Infectious Diseases, Medical Clinic, Research Center
Borstel, Borstel, GERMANY

Chi Chiu Leung

Tuberculosis & Chest Service, Department of Health, Hong Kong, CHINA

ABSTRACT

Currently, the standard short-course chemotherapy for tuberculosis comprises a six-month regimen, with a 4-drug intensive phase and a 2-drug continuation phase. Alternative chemotherapy using more costly and toxic drugs, often for prolonged durations generally more than 18 months, is required for multidrug-resistant and extensively drug-resistant tuberculosis. Directly observed treatment, as part of a holistic care programme, is a cost-effective strategy to ensure high treatment success and curtail development of drug resistance in tuberculosis. New antituberculosis drugs are urgently needed to improve the present standard short-course chemotherapy and alternative chemotherapy, by shortening administration durations and increasing cure rates, through the greater potency of these agents. At the same time, the role of adjunctive surgery for drug-resistant tuberculosis has to be better defined. Immunotherapy might improve treatment outcomes of both drug-susceptible and drug-resistant tuberculosis, and warrants further exploration.

INTRODUCTION

In the year 2008, 11.5 million people were estimated to be living with tuberculosis, with 9.4 million having incident disease. Among 1.9 million people who died of tuberculosis, 0.5 million were seropositive for HIV [1]. While the present chemotherapy for tuberculosis is highly efficacious, it has the disadvantages of being lengthy and complex, and does not live up to the expectation of adequately controlling the current global tuberculosis situation. In 2008, an estimated 390,000 – 510,000 cases of multidrug-resistant (MDR) tuberculosis with bacillary resistance to at least isoniazid (H) and rifampicin (R) are estimated to emerge every year worldwide, with China and India together accounting for approximately 50% of this global burden. In 2008, MDR tuberculosis caused an estimated 150,000 deaths [2]. Extensively drug-resistant (XDR) tuberculosis, recently emerging, is defined as MDR tuberculosis with additional bacillary resistance to any fluoroquinolone, and one or more of the three (second-line) injectable drugs – kanamycin, amikacin, and capreomycin. Approximately 5.4% of MDR tuberculosis reported worldwide could be categorized as XDR tuberculosis, with the proportion exceeding 10% in some countries [2]. This review examines the current status and future prospects of treatment of tuberculosis. Where appropriate, evidence levels for the recommended treatment regimens / modalities are given in accordance with the grading system of the Scottish Intercollegiate Guidelines Network (Appendix) [3].

Before discussing recommended drug regimens for treating of pulmonary tuberculosis, an understanding of basic mycobacteriology and antituberculosis drug action would be beneficial.

Scientific Basis of Short-Course Chemotherapy

Mycobacterium tuberculosis, the causative organism of tuberculosis, is a slowly growing bacterium, and it can also enter a phase of dormancy which appears drug-refractory. Four hypothetical populations of organisms [4] may exist in a patient with tuberculosis: 1) actively growing organisms, usually present in abundance (extracellularly) within aerated cavities, 2) slow intermittently growing organisms in an unstable part of the lesion, 3) organisms surviving under microaerobic conditions in a low environmental pH, either in inflammatory lesions or within phagolysosomes of macrophages, and 4) completely dormant organisms surviving under anaerobic conditions. The three major actions of antituberculosis drugs [5] are:

- bactericidal action, defined as their ability to kill actively growing bacilli rapidly, e.g. isoniazid, and to a lesser extent, rifampicin and streptomycin (S);
- sterilising action, defined as their capacity to kill the semi-dormant organisms, e.g. rifampicin and pyrazinamide (Z);

- prevention of emergence of bacillary resistance to drugs, e.g. isoniazid and rifampicin; less so for streptomycin, ethambutol (E) and pyrazinamide; least for thiacetazone and para-aminosalicylic acid.

CHEMOTHERAPY OF PULMONARY TUBERCULOSIS

Short-Course Chemotherapy Regimens

Based on a number of clinical trials performed previously, much knowledge has accumulated regarding chemotherapy regimens for new cases of smear-positive pulmonary tuberculosis [6-15]. The shortest duration of treatment required is at present six months (Grade A). The standard regimen today, as categorically recommended by the World Health Organization (WHO) / International Union Against Tuberculosis and Lung Disease (IUATLD) [16] comprises the combination of HRZE for two months, followed by that of HR for another four months. Streptomycin is not generally recommended as a fourth drug in the intensive phase, largely because of its higher resistance rate than that of ethambutol [17], and its requirement for parenteral route of administration. However, in rare occasions when ethambutol use is contraindicated, the aminoglycoside may be considered. Dosages for the conventional first-line antituberculosis drugs are well established, and can be found in standard references [16,18].

Although the 8-month regimen: 2 months of streptomycin, isoniazid, rifampicin and pyrazinamide, followed by 6 months of isoniazid and thiacetazone, combined with

hospitalisation in the first two months, has previously been shown to be effective in controlled clinical trials and programme settings in Africa [11], a randomized study initiated by IUATLD, has revealed that the 8-month regimen: 2HRZE / 6HE was significantly inferior to the 6-month regimen: 2HRZE / 4HR [19]. A systematic review has also shown that regimens utilizing rifampicin only for the first 1 – 2 months had significantly higher rates of failure, relapse and acquired drug resistance as compared to regimens that used rifampicin for 6 months [20]. The WHO currently recommends phasing out of the 8-month regimen [16]. Thus, short-course antituberculosis chemotherapy regimen with both rifampicin and pyrazinamide should contain six months, rather than two months, of rifampicin for better efficacy (Grade A).

A regimen without pyrazinamide in the initial intensive phase must be given for longer than six months (Grade A). Such regimen based on isoniazid and rifampicin [13-15] is only good for pansusceptible tuberculosis with limited bacillary load, and has to be given for nine months (namely 2HRE / 7HR or 9HR). This 9-month regimen is usually not recommended for patients in countries with high rates of isoniazid-resistant tuberculosis, except those who cannot tolerate pyrazinamide.

The administration of pyrazinamide beyond two months has not been shown to offer any advantage on treatment outcome (Grade A) [21,22]. Besides, in cohort and case-control

analyses, from 12 weeks or more after starting treatment, the estimated risk of hepatotoxicity was 2.6% for regimens incorporating pyrazinamide, isoniazid and/or rifampicin and 0.8% for standard regimens containing isoniazid and rifampicin. Thus adding pyrazinamide to isoniazid and rifampicin increases the risk of hepatotoxicity appreciably [23].

For individual cases with extensive disease and slow sputum bacteriological conversion, administration of pyrazinamide \pm ethambutol beyond two months may seem acceptable. This prolongation of intensive phase is not currently supported by WHO [16]. However, WHO has recently raised the possible advantage of using rifampicin, isoniazid and ethambutol rather than rifampicin and isoniazid in the continuation phase of treatment of tuberculosis in populations with known or suspected high levels of bacillary resistance to isoniazid [16]. Initial cavitation and positive sputum culture after two months of treatment have been found to be associated with increased risk of failure / relapse, and possibly justify prolongation of the continuation phase of antituberculosis therapy to give a total duration of nine months [24] (Grade B).

Intermittent regimens comprising two drugs in the continuation phase, following upon an intensive phase of four drugs given on a daily basis, have been proven to be highly efficacious (2HRZS / 4H₃R₃ or 2HRZS / 4H₂R₂) [7,10] (Grade A). The WHO does not

generally recommend twice-weekly regimens because of the higher risk of treatment failure when missing doses occur [16]. Intermittent short-course regimens administered thrice weekly all through have been shown to have largely equivalent efficacy to daily regimens [8]. A recent nested case-control study has raised concerns regarding the efficacy of three-times weekly 6-month regimens in preventing disease relapse in the presence of cavitation [25]. The systematic review just alluded [20] has not shown any significant difference in failure or relapse with daily or intermittent scheduling of treatment administration, apart from insufficient published evidence for the efficacy of twice-weekly rifampicin administration throughout therapy. However, major confounding factors, such as cavitation and 2-month culture status, might be heterogeneous across the included studies and not adequately controlled for in this systematic review. Furthermore, rates of acquired drug resistance among the failures / relapses have been shown to be higher with three-times weekly therapy [20]. Dosing schedules in the first nine weeks did not appear to have impact on the risk of hepatotoxicity in another case-control study [26]. Logistic regression analysis has shown that sex was nonsignificant but ageing increased the odds of hepatitis, the risk of which rose from 2.6% to 4.1% as age exceeded 49 years.

WHO currently recommends the use of daily dosing during both the intensive and continuation phase as the most optimal approach (Table 1).

HIV-infected patients who received 6-month rifampicin- or rifabutin-based regimen were shown to have a higher relapse rate than those on longer therapy in an early clinical trial [27] and a more recent treatment cohort [28]. Possibly because of the poor prognosis associated with the underlying HIV infection before the availability of antiretroviral therapy, the lower relapse rate did not translate into improved survival in the former trial [27]. WHO currently recommends that tuberculosis patients who are living with HIV should receive at least the same duration of treatment as HIV-negative patients. Increased risk of treatment failure and acquired rifamycin resistance has also been shown to be associated with intermittent regimens among HIV-infected patients [29-31]. WHO currently recommends that for HIV-positive patients with tuberculosis, and all tuberculosis patients living in HIV-prevalent settings should receive daily treatment, at least during the intensive phase [16].

In many countries, nearly 50% of patients are diagnosed as having active pulmonary tuberculosis on clinical and radiographic grounds, without immediate bacteriological confirmation. In the two smear-negative studies conducted in Hong Kong, it has been found that with 2 – 3 months of daily HRZS treatment, the relapse rates were 32% and 13% for culture-positive patients [32], but the rates were much lower with 4-month treatment (2% for

drug-susceptible tuberculosis and 8% for isoniazid- and streptomycin-resistant tuberculosis) [33].

Thus, it appears that at least 4 months of treatment is required for smear-negative pulmonary tuberculosis in non-HIV-infected patients (Grade C). The WHO currently recommends the use of a 6-month regimen of daily HRZE for 2 months followed by daily or three times per week HR for another 4 months in the treatment of new smear-negative pulmonary tuberculosis patients [16].

The relapse rates during 6 – 30 months after stopping the standard 6-month short-course chemotherapy regimen are generally less than 5% [6-10]. 78% of relapses occurred within 6 months of stopping treatment, and 91% within 12 months [34].

Retreatment Drug Regimens

For treatment of smear-positive relapse cases of pulmonary tuberculosis as well as retreatment after interruption, an 8-month regimen has been recommended by the WHO/IUATLD, namely 2HRZES / 1HRZE / 5HRE [16] (Grade D). With the increasing availability of rapid tests for bacillary drug susceptibilities, such as line probe assays, it would

be possible to modify this approach according to the results, particularly in areas with high prevalence of MDR / XDR tuberculosis [16]. Using conventional drug susceptibility testing, it might be necessary to start an empiric retreatment regimen active against MDR disease, when the levels of MDR tuberculosis are high in different patient registration groups in the geographical area (Grade D). Patients who have failed two rifampicin-containing regimens: the initial and retreatment ones are very likely to have MDR tuberculosis. These updated recommendations are now incorporated in the current WHO guidelines [16] (Table 2).

Directly Observed Treatment, Short-Course

Directly observed treatment (DOT) was shown to be highly efficacious in ensuring patient adherence by experience gained in Chennai (then Madras) and Hong Kong many decades ago. In 1993, the WHO officially announced the new global strategy for tuberculosis control known as directly observed treatment, short-course (DOTS) that implements the 6-month short-course regimen in a programmatic setting [35,36]. The DOTS strategy has five key components which include (i) a network of trained health-care or community workers to administer DOT, (ii) properly equipped laboratories with trained personnel to perform sputum microscopy for diagnosing tuberculosis, (iii) a reliable supply of high-quality drugs (preferably at no cost to patients), (iv) an accurate record keeping and cohort analysis system for

monitoring case-finding, treatment and outcomes and (v) sustained political commitment and funding. An effectively functioning tuberculosis control programme is clearly essential for good patient outcome [36]. Although some patient characteristics like homelessness, alcohol or substance abuse, behavioural problems, mental retardation, and lack of social or family support are more commonly associated with nonadherence to therapy, it is often difficult to identify poorly adherent patients because the underlying reasons for such behaviour are not only multifaceted and complex, but range from characteristics of the individual patients to qualities of the societal and economic environment [37]. Although a cochrane database systematic review concluded that the results of randomized controlled trials conducted in low-, middle-, and high-income countries did not provide assurance that DOT, compared with self-administered treatment, could impart quantitatively important effect on cure or treatment completion in tuberculosis patient [38], the great merit of reduction of acquired drug resistance with DOT was not addressed [39]. The DOTS strategy is more than DOT alone, it should be viewed as a comprehensive service, or an integral part thereof, which possesses ingredients also inclusive of enablers, incentives, education and holistic care that are conducive to the success of the treatment programme. In a cluster randomized controlled trial examining the effectiveness of a strategy to improve adherence to tuberculosis treatment in a resource-poor setting, the intervention package based on improved patient counseling and communication, decentralization of treatment delivery, patient choice of DOT supporter and reinforcement of

supervision activities led to improved patient outcome compared to the usual tuberculosis control procedures [40]. Apart from good communication skills of healthcare workers, attention regarding management of the treatment associated side-effects and risk of nonadherence, alongside maintaining respect for patient autonomy and integrity is of paramount importance [41]. One study has further demonstrated that both family-member and community DOTS strategies can attain international targets for treatment success under programme conditions [42].

Fixed-Dose Combination Formulations

The use of fixed-dose combination (FDC) formulations comprising 2 - 3 and even 4 drugs may enhance ease of prescription for physicians, reduce inadvertent medication errors, simplify drug procurement and supply, improve treatment adherence by patients, and thereby decrease the risk of development of MDR tuberculosis [43,44]. In a study that compared the levels of acquired drug resistance in patients who had rifampicin and isoniazid FDC, under self-administration settings, the rate was as low as 0.2%, given limitations of the investigation [45]. The WHO has included some FDC tablets in the list of essential drugs [46]. Only formulations of proven good quality should be used [47]. The majority of studies found no significant difference between FDC tablets and single drugs regarding sputum smear

conversion rates, side effects and relapses [48,49]. However, a Singapore study found higher relapse rates at 2 and 5 years of follow-up in patients who received FDC tablets [50]. Furthermore, FDC tablets cannot replace treatment supervision completely as there is still a potential risk of emergence of drug resistance when these combination tablets are taken irregularly [51].

Future Possibilities of Rifamycin Use

Studies have demonstrated the bactericidal and sterilizing activities of rifampicin, as well as their dose and concentration dependence [52-54]. In one study, the maximum dosage of rifampicin tested was 20 mg/kg [53]. Rifampicin at a dosage of >10 mg/kg may also suppress or delay emergence of resistance [54]. Early chemotherapy trials that evaluated the use of high-dose rifampicin have shown better 1-month or 2-month bacteriological conversion but not a more favourable relapse rate, likely due to absence of inclusion of pyrazinamide in the treatment regimens [55]. The safety and tolerability of high-dose rifampicin were not meticulously assessed in the early chemotherapy trials, thus leaving a potential concern for these issues.

Although rifampicin hepatotoxicity was thought to be idiosyncratic in nature [56], it is not possible to exclude interactive toxicity between isoniazid and rifampicin [57]. Thus it may not be entirely appropriate to extrapolate safety data from the use of high-dose rifampicin in treatment of other bacterial infections, such as brucellosis [55]. Mild hepatotoxicity in fact occurred more frequently in a study among patients who received high-dose rifampicin for tuberculosis treatment although no patient developed serious hepatotoxicity [58]. “Flu-like” syndrome has also been associated with high-dose rifampicin, but mainly for intermittent rifampicin administration, and it generally occurs after three months of drug administration [59]. Thrombocytopenia, haemolytic anaemia and acute renal failure may also occur. Since these reactions are immunological in origin, they are not likely to occur more frequently when a higher dosage of rifampicin is used [59]. Further clinical trials would be needed to examine whether such strategy could enhance bacillary sterilization and shorten tuberculosis therapy, without excessive adverse effects [60]. A phase II clinical trial is being conducted to compare the pharmacokinetics and pharmacodynamics of daily doses of 1200mg and 900mg of rifampicin with the standard 600mg dose during the two-month intensive phase of treatment [61].

Early trials of rifapentine, a long-acting cyclopentyl rifamycin, with a plasma half-life of 14 hours, given in a 600mg-dose together with isoniazid on a once-weekly basis during the

continuation phase of treatment in patients with tuberculosis have shown satisfactory efficacy in a subgroup of HIV-negative patients with noncavitary disease and limited bacterial burden, despite an overall failure/relapse rate of 10% [62,63], and emergence of rifamycin monoresistance in relapse cases among HIV-positive subjects [29]. It has been shown that a 900mg-dose of rifapentine had superior pharmacokinetics to the 600mg-dose [64,65], and that a 1200mg-dose of rifapentine produced an optimum pulse and postantibiotic lag on the growth of *M. tuberculosis* [52]. In a murine model, twice-weekly regimens containing rifapentine (15 – 20 mg/kg) have shown marked antituberculosis potency by enhancing the rifamycin exposure [66] and preliminary data using daily dosing of rifapentine have also been encouraging [67]. Rifapentine autoinduction of metabolism has also been noted with interest [68]. Currently there are only enough data on the safety and good tolerance of rifapentine dosed at 900 mg once weekly [69]. However, in an ongoing phase II clinical trial, rifapentine dosed at 10 mg/kg five days per week is being administered [70], and no unusual preponderance of adverse events was reported by the Data and Safety Monitoring Committee so far.

CHEMOTHERAPY OF PULMONARY MDR TUBERCULOSIS AND XDR TUBERCULOSIS

The clinical relevance of antituberculosis drug resistance is first reviewed as a background to the treatment of drug-resistant tuberculosis, especially the MDR and XDR forms.

Clinical Relevance of Antituberculosis Drug Resistance

Resistance to an antituberculosis drug arises spontaneously through chromosomal mutation at a frequency of 10^{-6} to 10^{-8} bacterial replications [36]. The chromosomal loci involved are distinct for the major classes of drugs. Thus, when three or more effective drugs are used in combination, spontaneous emergence of mutants resistant to all drugs is most unlikely with the usual bacterial load in the diseased host. However, sequential genetic mutations may be amplified through human error resulting in clinically drug-resistant tuberculosis. These include “monotherapy” due to irregular drug supply, inappropriate doctor prescription and, poor patient adherence to treatment [36]. Subsequent transmission of resistant *M. tuberculosis* strains from the index patient to others, as facilitated by diagnostic delay and infection-control breach, aggravates the problem [71]. A recent review regarding

epidemiology of MDR tuberculosis has shown that the risk factors for drug resistance pertain to those facilitating the selection of resistance in the community and the specific conditions that appear to increase the vulnerability of some patients, such as in certain HIV or malabsorption settings. The epidemiological situation is principally related to poor treatment practices and poor implementation of control programmes [71].

Isoniazid resistance is the most common form of drug resistance encountered, whether in isolation or in combination with other drugs [2]. Standard short-course chemotherapy for isoniazid-resistant tuberculosis can achieve a good success (>95% cure) when all four drugs are used throughout the six months of treatment (Grade B) [21]. When the four drugs are reduced to rifampicin and isoniazid after 2 months, the relapse rate after 6 months of treatment rises to 10% [22]. As there may be a genuine chance of resistance amplification with additional resistance to rifampicin [20,72], (especially for HIV status and/or intermittent dosing [20,28,31]), some authorities recommend changing to alternative regimens such as REZ, or RE, for more prolonged durations of administration, often up to one year (Grade D) [73,74]. Currently, the most optimal regimen for treatment of isoniazid-resistant tuberculosis appears unknown [16].

Rifampicin-resistant tuberculosis carries a much more ominous prognosis, as the outcome of standard short-course regimens for such disease is poor in terms of both disease status on cessation of treatment and subsequent relapse [75]. Recommendation has been made to treat such disease with EHZ for 18 – 24 months (**Grade D**) [76]. Some authorities feel that the duration of treatment can be shortened to 12 months by the addition of a fluoroquinolone to this 3-drug regimen (**Grade D**) [24]. Furthermore, rifampicin monoresistance in *M. tuberculosis* is usually rare, except perhaps in HIV-infected patients [28,31,76,77]. Thus, rifampicin resistance generally serves as a surrogate marker for dual resistance to rifampicin and isoniazid, i.e., MDR tuberculosis [78,79], especially for previously treated patients. Short-course chemotherapy can cure less than 60% of MDR tuberculosis cases [80], with a high recurrence rate of about 28% among those with apparent success [81]. A recent analysis has shown that the currently recommended short-course treatment regimens for both initial and retreatment purposes could not achieve good outcomes (with failures, relapses and deaths) in countries having initial MDR rates of greater than 3% [82]. It is quite clear today that alternative specific chemotherapy using second-line drugs is required for the management of this formidable condition [83].

Increased risk for development of bacillary resistance to ethambutol and pyrazinamide likely occurs when a conventional 4-drug regimen for initial treatment and a

conventional 5-drug retreatment regimen are repeatedly administered despite observed treatment failure with the conventional short-course regimens for tuberculosis [84-86]. Pyrazinamide and/or ethambutol resistance, in addition to dual resistance to isoniazid and rifampicin, generally portends a more adverse prognosis in MDR tuberculosis [87], particularly when patients receive only standardised second-line antituberculosis drug regimens with pyrazinamide and ethambutol plus a fluoroquinolone and aminoglycoside / capreomycin.

Fluoroquinolones are generally regarded as having a pivotal role in the treatment of MDR tuberculosis [88-90]. *In-vitro* resistance to fluoroquinolones has been shown to predict a poor outcome in the treatment of MDR tuberculosis [88,91,92]. Most fluoroquinolone resistance in *M. tuberculosis* is associated with the injudicious use of this class of drugs in the management of tuberculosis, particularly MDR tuberculosis [93,94], including the use of suboptimal second-line drug regimens comprising inadequate number and / or dosage / quality of accompanying agents [84]. Overzealous usage of this class of antimicrobials in the treatment of lower respiratory tract and other community-acquired infections might also contribute to development of fluoroquinolone-resistant tuberculosis [95].

As the aminoglycosides or capreomycin have potent antituberculosis activity, the loss of these second-line injectables together with fluoroquinolones through their suboptimal use in the

management of MDR tuberculosis would result in XDR tuberculosis, which in general carries a worse prognosis [96].

Programme Strategies and Implementation

The emergence of drug resistance in *M. tuberculosis* has prompted the WHO to modify the DOTS strategy to a more comprehensive approach – the Stop TB strategy [97]. This Strategy comprises the following components: (1) pursue high-quality DOTS expansion and enhancement, (2) address TB/HIV, MDR tuberculosis, and the needs of poor and vulnerable populations, (3) contribute to health system strengthening based on primary healthcare, (4) engage all care providers, (5) empower people with tuberculosis care, and communities through partnership and (6) enable and promote research. The management of MDR tuberculosis through use of alternative second-line antituberculosis chemotherapy mandates its delivery on a programmatic basis with five key components built on the DOTS framework (Figure 1). Capacity for performing drug susceptibility testing and availability of second-line drugs are not adequate to achieve cure. Other factors, such as a set of standard procedures, clear guidelines on treatment and follow up of patients and administration of DOT, must be included in the programme for MDR tuberculosis management to attain good results [83]. There are basically three possible programmatic approaches for the management of MDR tuberculosis [83]:

- Standardized treatment, in which regimens are designed on the basis of representative drug-resistance surveillance data of specific treatment categories, with all patients in the same group or category being treated by the same regimen.
- Empirical treatment, in which each patient's regimen is individually designed on the basis of the previous history of antituberculosis therapy with the help of representative drug-resistance surveillance data, followed by regimen adjustment when the individual drug susceptibility testing results are known.
- Individualized treatment, in which each patient regimen is designed on the basis of previous history of antituberculosis treatment and individual drug susceptibility testing results.

While a standardized regimen enables simple operation and broadens access to care, there may be a concern for amplification of multidrug resistance if the number of available second-line drugs in the regimen is low [98,99]. Individualized treatment strategies rely heavily on capable laboratory services, but have the advantage of avoiding placing patients on toxic and expensive drugs to which the *M. tuberculosis strain* is resistant. One caveat, however, is the unsatisfactory reliability of second-line drug susceptibility testing results, for many agents aside from the fluoroquinolones and the injectables, arising partly from the difficulty in standardizing testing methodology [100-102]. Some progress has, however, been made accordingly [103]. Patients who had previous treatment with second-line drugs would

likely benefit more from administration of individualized regimens. When retreated patients are presumed to have a high likelihood of MDR tuberculosis, an empiric regimen can be administered while awaiting results of conventional drug susceptibility testing [16].

Regardless of the strategy advocated, there are significant cost issues in the management of MDR tuberculosis patients [104]. From a previous decision analysis, more patients would die from tuberculosis, if the implementation of drug-resistance programme is associated with even minimal decreases in the effectiveness of DOTS [105]. Nevertheless, the feasibility and cost-effectiveness of treatment of MDR tuberculosis is now quite well established even in resource-limited settings [106,107]. In countries with significant financial difficulties, additional support, besides technical assistance, from international organizations and governments of industrialized countries would be needed, further to that obtained from local governments [108]. In this regard, the Green Light Committee of the Stop TB Partnership involving the WHO and collaborators has played a significant role in helping the implementation of these programmes in countries with an affordability problem in the management of MDR tuberculosis [109].

Design of Drug Regimens

Guidelines on the treatment of MDR tuberculosis are often formulated on experience and observational studies, as data from randomized trials are lacking. A detailed review has addressed the evidence and controversy of treatment of MDR tuberculosis, focusing on the number of antituberculosis drugs required to treat MDR tuberculosis, the most rational use of effective drugs against the disease, the advisable length of parenteral drug administration or of the initial phase of therapy, the contribution of surgery to the management of MDR tuberculosis and the optimal approach for treating MDR disease – standardized versus individualized regimens. However, little evidence but much controversy were found regarding the treatment of MDR tuberculosis [110]. Randomized controlled trials regarding chemotherapy of MDR tuberculosis should be undertaken to provide more evidence-based recommendations [111]. The updated WHO guidelines in 2008 and 2009 recommend designing treatment regimens with a consistent approach based on the hierarchy of five categories of antituberculosis drugs (Table 3) [16,112]. The potency of these drugs is in a descending order. Thus the drugs are so selected from these five groups accordingly. A brief review of the utility of these drugs is detailed below.

While isoniazid in conventional dose has limited usefulness, high-dose isoniazid (>10 mg/kg) has demonstrated some efficacy (clinical, bacteriological and radiographic) as well as reasonable patient tolerance in a recent study [113]. After adjustment for potential

confounders, subjects who received high-dose isoniazid had a 2.37 times higher likelihood of becoming culture negative at six months. Isoniazid-resistant *M. tuberculosis* organisms belonging to the low-resistance phenotype often have cross-resistance to ethionamide, while those of high-resistance phenotype are more susceptible. Adding high-dose isoniazid kills the former, leaving the latter that are more susceptible to ethionamide included in the MDR tuberculosis regimen. Ethambutol and pyrazinamide should be included in the treatment regimen if they are likely to be effective from laboratory evidence or clinical history.

Fluoroquinolone (in **Category 2**) therapy is independently associated with better treatment outcomes. Losing this drug category increases risk of death and failure [89,114]. Thus, a fluoroquinolone should be included whenever possible, although potential cross-resistance among the class members may hamper their utility in XDR tuberculosis [115]. Older fluoroquinolones, especially ciprofloxacin, are not recommended, as there could be slower sputum culture conversion and higher relapse rate [116]. Newer fluoroquinolones, moxifloxacin and levofloxacin, can be active against some ofloxacin-resistant strains of *M. tuberculosis* [117,118].

With the high rate of streptomycin resistance among MDR bacillary strains, an injectable agent from **Category 3**, capreomycin, kanamycin or amikacin, should form part of the regimen

as far as possible. Capreomycin may have a further advantage owing to its incomplete cross-resistance with kanamycin and amikacin in some *M. tuberculosis* strains [119,120]. However, variable cross-resistance exists among these three second-line injectables. Injectable agents are generally recommended for at least 6 months, or 4 months after culture conversion, with modification according to bacillary resistance or patient intolerance [83].

Category 4 agents are generally less efficacious and difficult to tolerate. Cycloserine / terizidone have potentially serious neurotoxicity. The use of thioamides and para-aminosalicylic acid are notoriously associated with gastrointestinal reactions and other adverse events [83,117]. They are added according to estimated bacillary susceptibility, drug history, efficacy, side-effect profile and cost.

Category 5 drugs, including linezolid, amoxicillin-clavulanate, imipenem / cilastatin and clofazimine, are not generally recommended in drug-resistant tuberculosis because their roles are uncertain [112]. However, they have potential role in situations without other options, especially in patients with XDR tuberculosis [16,121].

Clinical experience on linezolid has been slowly accumulating after the first report of its good *in vitro* activities against *M. tuberculosis* a decade ago [122]. In one study, 9 out 10

MDR tuberculosis patients given linezolid and other drugs under DOT setting were cured, despite substantial haematological and neurological toxicities [123]. Use of linezolid at half-dose (600 mg) daily helped to reduce bone marrow suppression, but not peripheral and optic neuropathy [124]. Fatal lactic acidosis can also occur after prolonged therapy [125]. A sizeable retrospective study has confirmed these adverse effects [126]. Most of them occurred after 60 days of therapy [126]. More major side-effects occurred with twice-daily than once-daily dosing, with no difference in efficacy. Outcomes were similar in patients treated with or without linezolid, although linezolid use was associated with more extensive resistance to first-line and second-line drugs. Thus it appears that linezolid 600 mg once daily, when added to an individualized multidrug regimen, may improve bacteriological conversion and treatment success in the most complicated MDR or XDR tuberculosis cases [126]. Its use might not be warranted where better tolerated alternatives are available. Further evaluation of linezolid at 600 mg daily dose is being conducted in a phase I/II clinical trial in South Africa [127]. Linezolid 300 mg daily, apart from safety, appears to have reasonable efficacy in a recent study [128]. However, concerns have been raised regarding the method of analysis and possible emergence of drug resistance [129]. For other oxazolidinones, PNU-100480, has demonstrated more potent activity *in vitro* and in the murine model [130], while AZD5847 is undergoing phase I trial in healthy volunteers [131].

Amoxicillin-clavulanate has some early bactericidal effect against *M. tuberculosis* [132] and distinct inhibitory activity on MDR strains [133]. Other β -lactam- β -lactamase inhibitor combinations have also shown similar *in vitro* activity [134], but clinical efficacy data are limited [135,136]. Imipenem / cilastatin is active against MDR and XDR strains *M. tuberculosis in vitro* [137]. There are also some limited efficacy data of imipenem in mice and humans [138]. When meropenem, a carbapenem – a newer congener of imipenem, was combined with the β -lactamase inhibitor clavulanate, potent activity against laboratory strains of *M. tuberculosis* was observed, with sterilisation of aerobically grown cultures achieved within 14 days [139]. In addition, this combination exhibited activity against anaerobically grown cultures that mimic the mycobacterial persisters, and inhibited the growth of 13 XDR strains of *M. tuberculosis* at the same levels as observed for drug-susceptible strains [139]. Thus, meropenem-clavulanate might have a potential role in the treatment of tuberculosis [140].

Clofazimine, a riminophenazine, and some of its analogues, have been shown to possess *in vitro* and *in vivo* activities against *M. tuberculosis*, including drug-resistant ones [141,142]. Clofazimine is primarily used in the treatment of leprosy but is sometimes incorporated in the treatment regimens for MDR tuberculosis, although data on its clinical efficacy [143] and

tolerance [144] are limited and conflicting. Rifabutin, has very limited potential utility in treatment of MDR tuberculosis due to its high cross-resistance rate with rifampicin [16].

MDR and XDR tuberculosis should be treated aggressively. The fluoroquinolones and injectables are the most potent second-line agents for MDR tuberculosis (Grade C). In the initial six months, the treatment regimen should consist of at least 4, non-cross-reacting drugs to which the organism is, or is likely to be, susceptible (Grade C). Generally speaking, those patients who have previously received second-line drugs are more likely to need a higher number of drugs. So do patients with more extensive radiographic disease and more formidable drug resistance patterns. The use of capreomycin/kanamycin/amikacin, fluoroquinolone, ethambutol, pyrazinamide, ethionamide/prothionamide for disease with bacillary resistance to RH (with/without S), and the use of capreomycin/kanamycin/amikacin, fluoroquinolone, ethionamide/ prothionamide, cycloserine, para-aminosalicylic acid for disease with bacillary resistance to RHEZ (with / without S) constitute important examples of these regimens. The possibility of further acquired resistance should be considered. A single drug should never be added to a failing regimen, for fear of selecting mutants that are resistant to the newly added drug (addition phenomenon) [84]. Care is also warranted in chemotherapy trials involving newly developed antituberculosis drugs. Linezolid resistance among *M. tuberculosis* is fairly well known today, and may serve as a warning [145,146].

The dosages of second-line drugs used in the treatment of MDR and XDR tuberculosis are listed in Table 4 [83,112,147]. The maximum dosage of cycloserine and ethionamide / prothionamide should generally be 750 mg when once-daily dosing is used, as there is concern with toxicity for higher doses. The same likely applies also for aminoglycosides / capreomycin given 3 – 5 times per week [147]. Some of the patients weighing >70 kg might tolerate 1000 mg for these three classes of drugs [112]. The maximum once-daily dosage of moxifloxacin and levofloxacin is 400 mg and 750 – 800 mg, respectively. While the efficacy of 1000 mg levofloxacin per day is high [148], the tolerance data are still limited.

The WHO recommends treatment duration of at least 18 months after culture conversion even for HIV-negative patients (Grade D) [83]. However, a proportion of immunocompetent patients who managed to achieve sustained sputum culture conversion early might be adequately treated with 12 months of fluoroquinolone-containing regimens [88,149]. It appears, however, that patients who are immunocompromised (including those with diabetes mellitus and silicosis), or have extensive radiographic evidence of disease (particularly with cavities), extensive drug resistance, delayed sputum culture conversion (i.e. after more than three months of chemotherapy) or extrapulmonary involvement should receive more than 12 months of therapy [88].

Another important principle in the chemotherapy of MDR tuberculosis / XDR tuberculosis is to exercise vigilance to prevent and manage adverse reactions.

Second-line drugs for treating MDR tuberculosis are generally more toxic and difficult to tolerate. In a study on MDR tuberculosis in Hong Kong, about 40% experienced adverse drug reactions of varying severity [88]. However, only half of them patients required modification of their drug regimens. These results corroborated the findings of a study of MDR tuberculosis patients in Peru where the adverse drug reactions never resulted in discontinuation of antituberculosis therapy, and only occasionally (11.7%) resulted in suspension of an agent [150]. In a reported series of MDR tuberculosis patients in Turkey, about 70% of them experienced adverse effects to the second-line agents, and 55.5% required treatment modification [151]. With timely and appropriate management, the treatment success rate (77.6%) did not appear to be compromised markedly. Indeed, the results from Turkey largely paralleled those pooled from five sites (Estonia, Latvia, Peru, the Philippines and the Russian Federation) in resource-limited settings [107].

The second-line antituberculosis drugs are handled in patients by different pathways, including diverse metabolic ones. There is a potential for them interacting with different classes of antiretroviral drugs [152].

Close clinical monitoring is necessary to ensure that adverse drug effects are recognized quickly. Apart from clinical monitoring, ancillary investigations, such as audiometry screen, vestibular assessment and biochemical tests including those of liver and renal functions, electrolytes and thyroid functions, are helpful. On top of assessment of visual acuity, tests to detect peripheral neuropathy are needed occasionally. The optimal intervals at which these investigations should be performed are unknown. Physicians should be aware, that some of the adverse effects that can occur during the continuation phase of an extended course of antituberculosis therapy can emerge within a few days. When an adverse reaction is mild and not dangerous, such as a gastrointestinal one, continuation of therapy alongside supportive treatment is sufficient. If an adverse event is severe or potentially dangerous, such as a neurological effect, a more intensive management strategy embracing supportive treatment and drug discontinuation or dosage adjustment is required [83]. Psychosocial support is also an important element in the management of adverse reactions. Education, counseling and encouragement can all contribute [83].

Patients developing nephrotoxicity were found to have a significantly longer duration of treatment with aminoglycosides and received a higher total dose. For ototoxicity of aminoglycosides, predisposing factors are less well characterized, except perhaps for old age, renal impairment and prolonged therapy [153]. Ototoxicity due to aminoglycosides can be irreversible and patients need to be counselled to report symptoms at the earliest signs of occurrence.

Treatment Outcomes

Treatment outcomes of MDR and XDR tuberculosis vary greatly between studies [88-90, 92,154-167], possibly related to variations in method of analysis, definition of treatment success and failure, drug susceptibility testing, clinical follow-up, and missing data. The outcomes also likely depend on adverse events due to drugs and their management, as well as supply / availability of the agents for treatment.

For MDR tuberculosis, success rates (cures + treatment completions) are around 50 – 70% [88-90,92,154-167]. In a recent systematic review [168], MDR tuberculosis treatment success rate improved with treatment duration of at least 18 months and DOT throughout treatment. Studies that combined both factors had significantly higher pooled success rate than other studies (69% vs 58%). Individualised treatment regimens conferred higher success (64%) than standardized regimens (54%) although the difference was not statistically significant. Patients with XDR tuberculosis in general had worse treatment success rates: $\leq 50\%$ [164-166], although a study has shown a remarkable treatment success rate of 60.4%, quite comparable with that (66.3%) of MDR tuberculosis in the same locality [169].

Prior antituberculosis therapy [89,90,92,154,157,167], extensive *in-vitro* drug resistance [87,89,92,157,165], fluoroquinolone resistance [88,90,92,158,167] or its prior use [158], capreomycin resistance [96], positive sputum smear [167], radiological cavitation [88,89,158], HIV seropositivity [156,161,163,166,167], other immunocompromised states [159], history of incarceration [92], low BMI ($<18.5/<20 \text{ kg.m}^{-2}$) [92,160], hypoalbuminaemia [164], older age [89,158], male sex [154], low haematocrit [160], and early (<1 year of treatment) default [163] constitute important risk factors for poor outcomes.

In a recent systematic review on XDR tuberculosis, it has been shown that strategies to support adherence, as well as psychological, nutritional and even financial interventions, might further contribute to improved outcomes in patients with XDR tuberculosis [170]. The encouraging results from some countries in Asia and Europe have also suggested that management in specialised reference centre would improve outcome, although high success rates could be achieved with treatment in some community settings [170].

The impact of HIV on the outcomes of MDR and XDR tuberculosis has been most serious in South Africa. Among 272 MDR and 382 XDR tuberculosis patients, with HIV-coinfection rates of 90% and 98%, one-year mortality was 71% and 83%, respectively. This mortality has improved, however, from 2005 to 2007, though the majority of death still occurred within the

first 30 days [166]. In the systematic review just alluded [170], it was remarked that additional data for HIV-infected individuals would be required to determine the role of HIV coinfection in XDR tuberculosis treatment outcome, and to evaluate interventions that might contribute to improve outcomes in HIV-infected XDR tuberculosis. The high case-fatality rate in the Tugela Ferry outbreak could represent a combination of factors at play – not only host immunocompromisation, but also lack of access to adequate diagnosis and treatment [170].

Palliative Management and End-of-Life Care

At a certain time-point, recourse to palliative care is indicated for selected patients with “difficult” MDR or XDR tuberculosis, in the interest of both the individual patient and the community as a whole. Such management aims to provide uninterrupted medical and psychological care, as well as to ensure a dignified termination of the patient [83,147].

NEW DRUGS FOR TREATMENT OF TUBERCULOSIS

New antituberculosis drugs are needed to simplify treatment of drug-susceptible tuberculosis and to improve outcome of drug-resistant tuberculosis [171]. Only four compounds would be discussed in some depth for the purpose of this review, as these drugs appear to have sufficient potential for utilization to improve tuberculosis therapy in the coming decade.

Moxifloxacin

Moxifloxacin is a 8-methoxy fluoroquinolone with a long plasma half-life of approximately 11 hours. It has potent bactericidal and sterilizing activity against *M. tuberculosis*, as shown in murine studies [172,173]. In the TB Trials Consortium (TBTC) Study 27, a Phase II trial, substituting moxifloxacin for ethambutol in the first 8 weeks of therapy did not change the 2-month culture negativity rates (71%), but there appeared to be higher activity at earlier time point [174]. On the other hand, a study with nearly the same design conducted in Brazil has shown better culture conversion (80%) in patients receiving moxifloxacin during the initial phase of treatment, compared with the ethambutol arm (63%) [175]. Another similar Phase II clinical study has also shown that patients in moxifloxacin -containing arm

cleared their sputum bacilli more quickly [176]. Based on the rapid sterilization results of isoniazid-sparing regimen in murine models, the TBTC Study 28 was designed as a double-blind, placebo-controlled study to evaluate the 2-month culture conversion rates with the substitution of moxifloxacin for isoniazid in the 2-month intensive phase of treatment of pulmonary tuberculosis. Only a small nonsignificant increase in the 2-month culture negativity was achieved [177]. More rapid sputum culture conversion was also observed with the addition of moxifloxacin to the standard short-course regimen in a non-randomized study [178]. In these studies, moxifloxacin appeared to be well tolerated by most patients, apart from an increased incidence of nausea. QTc prolongation was observed in some patients but might not have clinical significance [175,177]. Further evaluation of the fluoroquinolone is ongoing in a phase III REMox study [179]. This study will explore whether moxifloxacin substitution for isoniazid or ethambutol can shorten the conventional therapy from 6 months to 4 months.

Moxifloxacin and rifapentine based regimens are also under investigation. It should be noted that when given together, rifapentine may induce enzymes that metabolize moxifloxacin, resulting in modestly reduced moxifloxacin concentrations [68]. Using a murine model of tuberculosis, regimens consisting of isoniazid or moxifloxacin plus rifapentine and pyrazinamide administered either daily or three-times weekly were evaluated for bactericidal

activity and treatment-shortening potential. The duration of treatment necessary to achieve stable cure was 10 weeks for daily regimens and 12 weeks for thrice-weekly regimens, regardless of whether isoniazid or moxifloxacin was used with rifapentine and pyrazinamide [180]. By contrast, for the 12-week regimen of RHZ, all mice relapsed. The treatment-shortening potential of more frequent and /or higher doses of rifapentine than 600 mg once weekly are being explored in both animal experiments and clinical trials, as discussed in the section on treatment of smear-positive pulmonary tuberculosis. Furthermore, in a Phase II clinical trial that commenced in 2009, smear-positive pulmonary tuberculosis patients are being randomized in the initial 2 months to receive either H-rifapentine-Z-moxifloxacin or RHEZ, followed by the standard nonexperimental regimen in the continuation phase. The efficacy in terms of sputum conversion rates and treatment outcomes, as well as safety and tolerability of the rifapentine-moxifloxacin-containing regimens, will thus be evaluated [181].

Notwithstanding these somewhat encouraging results, the high rates of fluoroquinolone-resistant tuberculosis in many parts of the world, especially coinciding with high disease burden, poses concern regarding the potential utility of the new fluoroquinolones in shortening tuberculosis treatment [182]. Regarding the role of moxifloxacin in the treatment of MDR tuberculosis / XDR tuberculosis, there has been some promising results lately [183], although the issue of partial cross-resistance among *M. tuberculosis* strains still casts concern [184].

TMC207

TMC207 is a novel diarylquinoline with unique activity on the mycobacterial ATP synthase [185]. It is active against both drug-resistant and drug-susceptible *M. tuberculosis*, as well as other mycobacterial species. It has a long half-life in plasma and tissues of nearly 24 hours. Data have also suggested that TMC207 might kill dormant bacilli as effectively as aerobically grown bacilli. TMC207 is metabolized by CYP3A4, thus, its plasma level may be reduced by half through interaction with rifampicin. However, data from the mouse model have demonstrated that TMC207 had significant activity, even when its exposure was reduced by 50% and when it was added to a strong background regimen of RHZ. The bactericidal effect of TMC207 in mice was modest during the first week of treatment but increased in the following three weeks [186]. TMC207 probably acts synergistically with pyrazinamide to exert sterilization activity. In the mouse model, 2-month treatment regimens containing TMC207 and pyrazinamide led to sterilization, suggesting treatment-shortening potential [187]. In another mouse model study, the triple combination of TMC207-rifapentine-pyrazinamide given once weekly was more active than the current regimen of RHZ given five-times per week and led to satisfactory lung culture negativity at two

months [188]. Such unprecedented activity has suggested that it might be feasible to develop a fully intermittent, once-weekly regimen.

In a mouse model to evaluate the use of TMC207 in MDR tuberculosis, treatment was given five times per week with TMC207 alone or various combinations of TMC207 plus pyrazinamide or other second-line drugs [189]. All TMC207-containing regimens were significantly more active than the non-TMC207-containing regimens after one month of therapy.

An early bactericidal activity study with ascending doses of TMC207 has demonstrated a delayed onset of bacteriolysis, with significant activity from day 4 – 7 when given at a daily dose of 400 mg, which was similar in magnitude to those of isoniazid and rifampicin over the same period [190].

A double-blind, randomized Phase II clinical trial with TMC207 began in 2007 in MDR tuberculosis patients [191]. The study is being conducted in two consecutive stages. In the first exploratory stage for safety and dose determination, newly diagnosed sputum smear-positive patients with MDR tuberculosis were randomized to receive either TMC207 or placebo for eight weeks on top of a background regimen. The dosing scheme for TMC207

was validated for further testing in the second stage, being 400 mg daily for two weeks followed by 200 mg three-times weekly. In the second stage planned for proof of effectiveness, patients are randomized to receive either TMC207 or placebo for 24 weeks on top of a background regimen. After finishing 24 weeks of treatment, patients will continue to receive MDR tuberculosis treatment as per national treatment guidelines. Study subjects will be followed for safety, tolerability, pharmacokinetics, and microbiological efficacy for 96 weeks after receiving their last dose of TMC207 [191]. Preliminary results in the first stage indicated high efficacy (faster rate and higher proportion of culture conversion) and good tolerance of TMC207 [192]. Further results are awaited with great interest. Like moxifloxacin, QTc prolongation with uncertain clinical significance was observed in some patients, aside from gastrointestinal upset [192]. TMC207 can have a role in treatment of MDR tuberculosis and XDR tuberculosis, subjected to confirmation of its tolerance and safety on long-term use. Its interaction with rifampicin might hamper its utility in treatment of drug-susceptible tuberculosis.

OPC-67683

OPC-67683 is a nitroimidazole with high potency *in vitro* and *in vivo* against *M. tuberculosis*, inclusive of MDR strains. It probably acts through inhibition of cell wall

biosynthesis, a mechanism similar to that of PA-824 [193], but is approximately 20-times more potent. OPC-67683 and PA-824 are closely related compounds and appear to show cross-resistance. OPC-67683 has a long half-life at approximately 7 – 8 hours, with no cross-resistance or antagonistic activity with first-line antituberculosis drugs. In addition, it has promising post-antibiotic effects against *M. tuberculosis* intracellularly, comparable to that of rifampicin [193]. In mice, 2 months of OPC-rifampicin-pyrazinamide followed by another 2 months of OPC-rifampicin led to complete culture negativity, suggesting that OPC-67683 in combination with other existing drugs could potentially shorten tuberculosis therapy [193]. A randomized, double-blind, multicentre Phase II clinical trial has been underway since 2008 to evaluate its safety, efficacy and pharmacokinetics in the treatment of MDR tuberculosis. In the first 56 days, patients receiving an optimized background regimen were randomized to receive either placebo or OPC-67683 at a dose of 100 or 200 mg twice daily. Thereafter, the study subjects will complete their optimized background regimen [194].

PA-824

PA-824 is a nitroimidazopyran, a class of novel antibacterial agents. It is active against drug-susceptible and drug-resistant, and both dividing and nonreplicating *M. tuberculosis* [195]. From studying colony-forming-unit counts in the lungs of mice, PA-824 showed

bactericidal activity comparable to that of isoniazid in the first eight weeks and sterilizing activity comparable to that of HR in the continuation phase [196]. A follow-up experiment in mice showed advantages in relapse rate with the same combination of drugs when PA-824 was given at a higher dose of 100 mg/kg [197]. With the novel combination of PA-824-moxifloxacin-pyrazinamide, mice were cured more rapidly than with the first-line regimen of RHZ, suggesting that this combination might radically shorten the treatment of MDR tuberculosis in humans [198].

No serious adverse events were reported in Phase I single-dose and multiple-dose trials in healthy volunteers [199]. An extended early bactericidal activity study was conducted recently in South Africa, evaluating the efficacy, safety and pharmacokinetics in newly diagnosed sputum smear-positive patients with drug-susceptible tuberculosis [200]. Based on findings from the preclinical and Phase I studies, escalating doses of PA-824 were administered for 14 consecutive days for four groups of patients and compared with a fifth cohort receiving standard first-line antituberculosis treatment. The study showed substantial and continued early bactericidal activity over 14 days with equivalent efficacy at all doses from a daily dose of 200 – 1200 mg. One important feature of PA-824 is its high protein binding (94%). Thus, it is necessary to ensure that sufficiently high concentrations of the free drug can be reached in cavities of pulmonary tuberculosis to exert bactericidal activity [201]. Both OPC-67683 and

PA-824 appear to have potential roles in treatment of drug-susceptible and drug-resistant tuberculosis. Again, the most critical determinant would be their safety profiles.

Other Potential Candidates

Important examples of two such compounds include a pyrrole derivative LL3858 and a diamine compound SQ109. Both have been subjected to Phase I testing and further progress is ongoing [55,202,203]. Other potential candidates would be those have both potent bactericidal and sterilizing activities. Examples might include ATP synthase inhibitors, gyrase inhibitors and peptide deformylase inhibitors [203]. Aside from these new drugs under development, a neuroleptic thioridazine with impressive antituberculosis activity might also warrant repurposing to constitute a new agent for treating MDR tuberculosis and XDR tuberculosis [204].

ADJUNCTIVE SURGERY FOR PULMONARY TUBERCULOSIS

While chemotherapy using antituberculosis drugs constitutes the primary treatment modality for pulmonary tuberculosis, emergence of MDR and XDR tuberculosis has rekindled the enthusiasm in recourse to adjunctive surgery to improve the chance of cure in some patients in these drug-resistant scenarios [205]. Other indications of surgical treatment of tuberculosis centre on management of empyema, post-tuberculous bronchiectasis and mycetoma [206].

There are three basic selection criteria for adjunctive surgery in MDR tuberculosis patients [205]. These include:

- drug resistance, as revealed by *in vitro* susceptibility testing, is so severe or extensive that there is a high probability of failure or relapse with medical therapy alone
- disease is sufficiently localized that the great preponderance of radiographically discernible disease can be resected with expectation of adequate cardiopulmonary capacity post-surgery
- drug activity is sufficient to diminish the mycobacterial burden to facilitate healing of bronchial stump after lung resection

Patients should receive chemotherapy prior to surgery for at least 3 months [205,207]. If possible, they should be rendered culture negative before lung resection. However, this may not always occur. In some cases, sputum culture conversion only appears with prolonged medical therapy after surgery. Ventilation / perfusion scan, pulmonary function tests and computed tomography of the chest are important investigations for pre-operative assessment [205,206]. For some patients, assessment of pulmonary artery pressure and bronchial tree anatomy / pathology needs to be performed. Bilateral disease does not necessarily preclude surgical intervention, unless extensive [208]. Such disease would, however, require staged bilateral resection. In experienced hands, the outcome of lung resection has been found to be rather rewarding. The cure rates could reach $\geq 90\%$ with post-surgery chemotherapy [209-222]. (**Table 5**) In resource-limited areas, the cure rates might be lower (63 – 75%), but lung resection still appears useful as adjunctive management for this formidable disease [218]. However, adjunctive surgery necessitates expertise and financial instillation which are often not readily available in many areas where MDR tuberculosis prevail [223].

Two rather sizable cohort studies have shown that the best outcomes of MDR tuberculosis patients were achieved by the use of fluoroquinolones and adjunctive surgery [89, 224]. Although there has been no randomized study to compare chemotherapy alone versus combined chemotherapy and surgery in the management of MDR tuberculosis, one recent

small series reported significant and durable improvement with lung resection and post-operative first-line antituberculosis chemotherapy in patients with MDR and XDR tuberculosis [225]. This finding suggests the possibility of an independent role of lung resection in the management of these difficult drug-resistant scenarios.

With the emergence of XDR tuberculosis, adjunctive surgery becomes more relevant [121,225]. Notwithstanding its use in some patients, the outcome of XDR tuberculosis is generally worse than that of MDR tuberculosis, with an overall success rate of about 50% [121,227]. However, in selected patients, sustained sputum bacteriological conversion can be satisfactorily maintained [228]. From two large series, patients who had adjuvant surgery experienced better outcomes [89,229].

Regarding the factors governing outcome of surgery for drug-resistant tuberculosis, a low body mass index ($<18.5 \text{ kg/m}^2$), bacillary resistance to fluoroquinolones and presence of cavity beyond the range of surgical resection portended poor prognosis in a carefully performed study [219]. The major complications of surgical treatment of pleuro-pulmonary tuberculosis include broncho-pleural fistula, residual space problem and empyema. Other complications include wound and other infections, bleeding, cardiovascular embarrassment, atelectasis, and recurrent laryngeal nerve injury. The risk factors for broncho-pleural fistula mainly include

sputum-smear positivity, low FEV₁, old age, and perhaps the technique of stump closure and reinforcement [230]. There have been no randomized controlled studies that compared bronchial stump reinforcement versus nothing or stapling versus suturing as a means of closure of the stump. Many authorities, however, have recommended reinforcement of the bronchial stump, especially in selected patients at risk of such complications [222,230].

In some frail patients with MDR tuberculosis, who usually have limited cardiopulmonary reserve and thus would not withstand lung resection, collapse therapy using thoracoplasty [231], plombage [232] and artificial pneumothorax [233] can be considered. Thoracoplasty, aside from causing cosmetically unappealing deformity of the thoracic cage, can be associated with obstructive and restrictive lung function defects after the procedure. Extra-pleural lucite sphere plombage can give rise to pressure effects, migration, and foreign-body irritation problems. The use of artificial pneumothorax has been reappraised in a recent study with rather encouraging radiographic and bacteriological results [233]. In the intervention group, culture negativity was achieved in all new cases and 81.1% retreatment cases (About 80% patients had MDR tuberculosis). Cavity closure occurred in 94.6% and 67.9%, respectively. In the control group, culture negativity was achieved in 70.9% and 40.0%, respectively, and cavity closure occurred in 56.3% and 24.0% respectively.

ADJUNCTIVE IMMUNOTHERAPY IN TUBERCULOSIS

Macrophages, dendritic cells, natural killer cells, $\gamma\delta$ T cells and CD1-restricted T cells are involved in the initial cell-mediated immune response to *M. tuberculosis*, and determine the local / distant progression of infection to disease, versus containment of the infection. Antigens of *M. tuberculosis* are processed by the antigen-presenting cells. Subsequently CD4⁺ cells are involved. T helper (Th) cells, largely CD4⁺ cells, generally mature into two functionally different phenotypes, often termed Th1 and Th2 cells. The former secrete principally interleukin-2 and interferon- γ , while the latter largely secrete or induce interleukin-4, interleukin-5, interleukin-6 and interleukin-10. Interleukin-12 produced by macrophages, expands Th1 cell population and upregulates its functions. Cell-mediated protective immunity appears to be associated with a Th1 response [234]. Interleukin-18, another cytokine linked to Th1 pathway, may also have a putative role in cell-mediated protection against mycobacterial infection [235]. Tumour necrosis factor alpha (TNF- α), released largely from macrophages, contributes to protect the host by promoting granuloma formation [236]. However, TNF- α can also cause tissue necrosis under subversive T-cell influence. There is some evidence this sabotage effect comes from interleukin-4 overactivity [237]. **Figure 2** summarizes the immunopathogenesis of tuberculosis. Thus, the complex

immunopathogenesis of tuberculosis embraces host tissue inflammation and damage, on top of protective immunity against the tubercle bacilli.

Although a recent in-depth review on the immunotherapy for tuberculosis has revealed a number of potentially useful agents for immuno-regulation, immuno-augmentation or immuno-suppression, no evidence-based recommendation can yet be formulated regarding their clinical utility [238].

Adjunctive corticosteroids have been used as an attempt to ameliorate the inflammation. Cochrane reviews have shown improved mortality of patients with tuberculous pericarditis [239] and tuberculous meningitis [240] with steroid therapy, but inconclusive effects for pericardial constriction. Neurological deficit / disability was improved among survivors with tuberculous meningitis. There is currently inadequate evidence on whether steroids are effective in tuberculous pleural effusion [241]. In HIV-infected patients, steroids have been shown to be beneficial in tuberculous meningitis, although the overall prognosis is still poor [242]. Steroid use in tuberculous pleural effusion in HIV-infected patients was associated with a higher incidence of Kaposi's sarcoma [243]. An additional concern is that adjuvant steroid therapy of HIV-related tuberculosis has been associated with a transient increase in HIV

viral load [244]. In these two latter studies, the dosages of prednisolone used were 50 mg/day and 2.75 mg/kg/day, respectively, with gradual tapering off in eight weeks [243,244].

Cytokine supplementation was initially thought to be promising adjunctive therapy in tuberculosis [245], including drug-resistant forms. Table 6 depicts the results of a number of preliminary studies regarding the use of cytokines especially interferon- γ) [246-251] and *Mycobacterium vaccae* (NCTC 11659) [252], an avirulent vaccine from a nontuberculous mycobacterial species, in the management of MDR tuberculosis. In a more recent study, nebulised interferon- γ 1b adjuvant therapy has also been found to improve constitutional symptoms, reduce inflammatory cytokines in bronchoalveolar lavage, and improve clearance of acid-fast bacilli from sputum in cavitary pulmonary tuberculosis [253]. While the results from some of these studies are encouraging, the limited number of enrolled patients, alongside often uncontrolled experimental designs, leaves great uncertainty regarding the definitive role of these cytokines and allied forms of immunotherapy in tuberculosis treatment.

By enhancing mycobacterial killing in macrophages, vitamin D might have the potential to enable shortening of treatment duration for tuberculosis, reducing infectiousness, and improving response in drug-resistant forms of the disease [254]. However, a double-blind, randomized, placebo-controlled trial [255] has recently shown that vitamin D, as

supplementary therapy, did not improve clinical outcome (as assessed by clinical score severity and sputum smear conversion) among patients with tuberculosis. There was also no overall effect on tuberculosis mortality at 12 months. One caveat might be the possibly insufficient dose of vitamin D used. The clinical role of vitamin D in immunotherapy of tuberculosis is currently uncertain.

In addition, there appears to be some agents that can promote intracellular killing of *M. tuberculosis* by macrophages, through affecting the transport of K^+ and Ca^{2+} from the phagolysosome, thereby resulting in better acidification and activation of hydrolases [256]. This might be a promising direction of developing therapy for drug-resistant tuberculosis.

CONCLUSION

In 2010, the prevailing challenges of HIV infection and drug resistance still undermine the global control of tuberculosis. With clear indications that XDR tuberculosis results from mismanaged cases of drug-susceptible and MDR tuberculosis, it would be imperative to treat drug-susceptible tuberculosis appropriately to completion, and to provide rapid diagnosis, and aggressive as well as appropriate treatment of MDR tuberculosis to avoid the unnecessary development of additional cases of XDR tuberculosis. The main priority interventions would be (1) strengthening control of tuberculosis (through sound implementation of the Stop TB Strategy, with special focus on laboratory capacities and infection control (including HIV control) (2) improvement of programmatic management of drug-resistant tuberculosis, based on updated guidelines, largely from WHO, and (3) promotion of research and development of new diagnostics, vaccines and drugs, as well as other modalities of therapy.

While scientific advancement is crucial to better the care of tuberculosis patients and is earnestly awaited, existing tools must be harnessed in sound public health settings to curb the epidemic of tuberculosis today [257].

Table 1 Recommended Dosing Frequency for Standard Six-Month Regimen (Adapted with permission from WHO/HTM/TB/2009.420)

Dosing frequency		Comments
Intensive phase	Continuation phase	
Daily	Daily	Optimal
Daily	3 times per week	Acceptable alternative for any new tuberculosis patient receiving directly observed therapy
3 times per week	3 times per week	Acceptable alternative provided that the patient is receiving directly observed therapy and is NOT living with HIV or living in an HIV-prevalent setting

Table 2 Suggested Antituberculosis Retreatment Regimens for Previously Treated Patients (Adapted with permission from WHO/HTM/TB/2009.420)

DST	Likelihood of MDR-TB	
Routinely available	High (Failure patients)	Medium/Low (Relapse/Default patients)
Rapid molecular tests	DST results available in 1 – 2 days to confirm or exclude MDR-TB to guide treatment regimen used	
Conventional phenotypic tests	While awaiting DST results Empiric MDR-TB regimen (to be modified once DST results are available)*	While awaiting DST results 2HRZES/HRZE/5HRE (to be modified once DST results are available)*

DST = Drug susceptibility testing

MDR-TB = Multidrug-resistant tuberculosis

*Standardized / Individualized regimen if MDR-TB is confirmed

Table 3 Categories of Antituberculosis Drugs (Adapted with permission from WHO/HTM/TB/2009.420)

Category 1: First-line oral drugs

Isoniazid
Rifampicin
Ethambutol
Pyrazinamide

Category 2: Fluoroquinolones

Levofloxacin
Moxifloxacin
Ofloxacin

Category 3: Injectable agents

Capreomycin
Amikacin
Kanamycin
Streptomycin

Category 4: Oral bacteriostatic second-line agents

Ethionamide
Prothionamide
Para-aminosalicylic acid
Cycloserine
Terizidone

Category 5: Agents with efficacy that is not totally clear / certain (not recommended by the WHO for routine use in treating patients with drug-resistant tuberculosis generally)

Isoniazid (high-dose: >10 mg/kg)
Linezolid
Amoxicillin-clavulanate
Clarithromycin
Clofazimine
Imipenem / cilastatin (+ clavulanate)
Thiacetazone
Rifabutin

Table 4 Dosages of Antituberculosis Drugs Used in Treatment of MDR tuberculosis

in Adults

Group	Drug	Daily dosage/kg [†]	Usual daily dosage [†]
Aminoglycosides + allied injectable peptide antibiotics	Streptomycin	15 mg/kg	750 mg [‡]
	Kanamycin	15 mg/kg	750 mg [‡]
	Amikacin	15 mg/kg	750 mg [‡]
	Capreomycin	15 mg/kg	750 mg [‡]
Fluoroquinolones	Ofloxacin	-	600 – 800 mg [‡]
	Levofloxacin	-	500 – 750 mg ^{‡¶}
	Moxifloxacin ^{‡‡}		^{‡‡} 400 mg [‡]
Thioamides	Ethionamide [§]	15 mg/kg	500 – 750 mg [‡]
	Prothionamide [§]	15 mg/kg	500 – 750 mg [‡]
Oral bacteriostatic first-line drugs	Ethambutol	15 mg/kg	-
	Pyrazinamide	20 – 30 mg/kg	1 – 2 g [‡]
Oral bacteriostatic second-line drugs	Cycloserine [§]	15 mg/kg	500 – 750 mg [‡]
	Terizidone [§]	15 mg/kg	600 mg [‡]
	Para-aminosalicylic acid ^{§§}	0.2 g/kg	8 – 12 g [‡]
	Thiacetazone ^{¶¶}	2.5 mg/kg	150 mg [‡]
Oral reserve drugs with uncertain / not totally clear antituberculosis activities	Clofazimine	-	^{‡‡} 100 mg [‡]
	Amoxicillin-clavulanate ^{§§††}	-	^{‡‡} 875 – 125 mg 2X/day [‡] or
		-	^{‡‡} 500 – 250 mg 3X/day [‡]
	Linezolid ^{§§††}		^{‡‡} 600 mg 2X/day [‡] or ^{‡‡} 600 mg

[†] Drugs are generally given on a daily basis except for aminoglycosides and the allied injectable antibiotics, which are given 3X to 5X/week, as well as those otherwise specified

[‡] Usually the maximum daily dosage (Some of the patients weighing >70 kg can tolerate higher dosages – See text)

[§] May require administration in 2 split doses per day

[¶] Higher dosage usually given for fluoroquinolone-resistant disease

^{††} Long-term safety not fully confirmed

^{‡‡} Optimal dosage not fully delineated

^{§§} Requires administration in split doses per day

^{¶¶} Should only be used in patients documented to be HIV-negative, and usually not be chosen over other oral bacteriostatic second-line drugs

Table 5 Surgical Treatment of MDR Tuberculosis*

Investigator	Patient number	Treatment success rate (%)	Operative mortality rate (%)	Postoperative complication rate (%)
Treasure <i>et al</i> [209]	19	89	0	9
van Leuven <i>et al</i> [210]	62	75	2	23
Sung <i>et al</i> [211]	27	96	0	26
Pomerantz <i>et al</i> [212]	172	98	3	12
Chiang <i>et al</i> [213]	27	92	4	11
Park <i>et al</i> [214]	49	94	0	16
Naidoo <i>et al</i> [215]	23	96	0	17
Takeda <i>et al</i> [216]	26	89	3	14
Kir <i>et al</i> [217]	79	95	3	5
Somocurcio <i>et al</i> [218]	121	63	5	23
Kim <i>et al</i> [219]	79	72	1	23
Mohsen <i>et al</i> [220]	23	96	4	35
Wang <i>et al</i> [221]	56	87	0	25
Shiraishi <i>et al</i> [222]	56	98	0	16

** Most patients had lung resections in form of pneumonectomy or lobectomy (A minority also had segmentectomy).*

Table 6 Preliminary Studies on Adjunctive Immunotherapy of MDR Tuberculosis

Investigator	Immunomodulator/ Cytokine	Outcome
Condos <i>et al</i> [246]	IFN- γ (aerosolised)	Bacillary load lowering, radiographic improvement (CT)
Johnson <i>et al</i> [247]	rhIL-2 (subcutaneous)	Reduced bacillary load, radiographic improvement
Palmero <i>et al</i> [248]	r-IFN- α 2b (subcutaneous)	Bacillary load reduction
Giosue <i>et al</i> [249]	IFN- α (aerosolised)	Bacillary load reduction, radiographic improvement (CT)
Grahmann <i>et al</i> [250]	IFN- γ (aerosolised)	Bacillary load lowering, radiographic improvement
Park <i>et al</i> [251]	IFN- γ (subcutaneous)	No bacteriological conversion on smear and culture, no radiographic improvement (CT)
Stanford <i>et al</i> [252]	<i>Mycobacterium vaccae</i> (intradermal)	<u>Disease ≤ 2 yr:</u> Cure rate: 82% (1 to 2 doses) <u>Chronic cases:</u> Cure rates: 7.6% (1 dose) 37.9% (7 doses) 41.6% (12 doses)

CT computed tomograph
 IFN- α interferon-alpha
 IFN- γ interferon-gamma
 rhIL-2 recombinant human interleukin-2
 rIFN- α 2b recombinant interferon-alpha2b

Figure 1 DOTS Framework Applied to the Management of Drug-Resistant Tuberculosis (Adapted with permission from WHO/HTM/TB/2006.361)

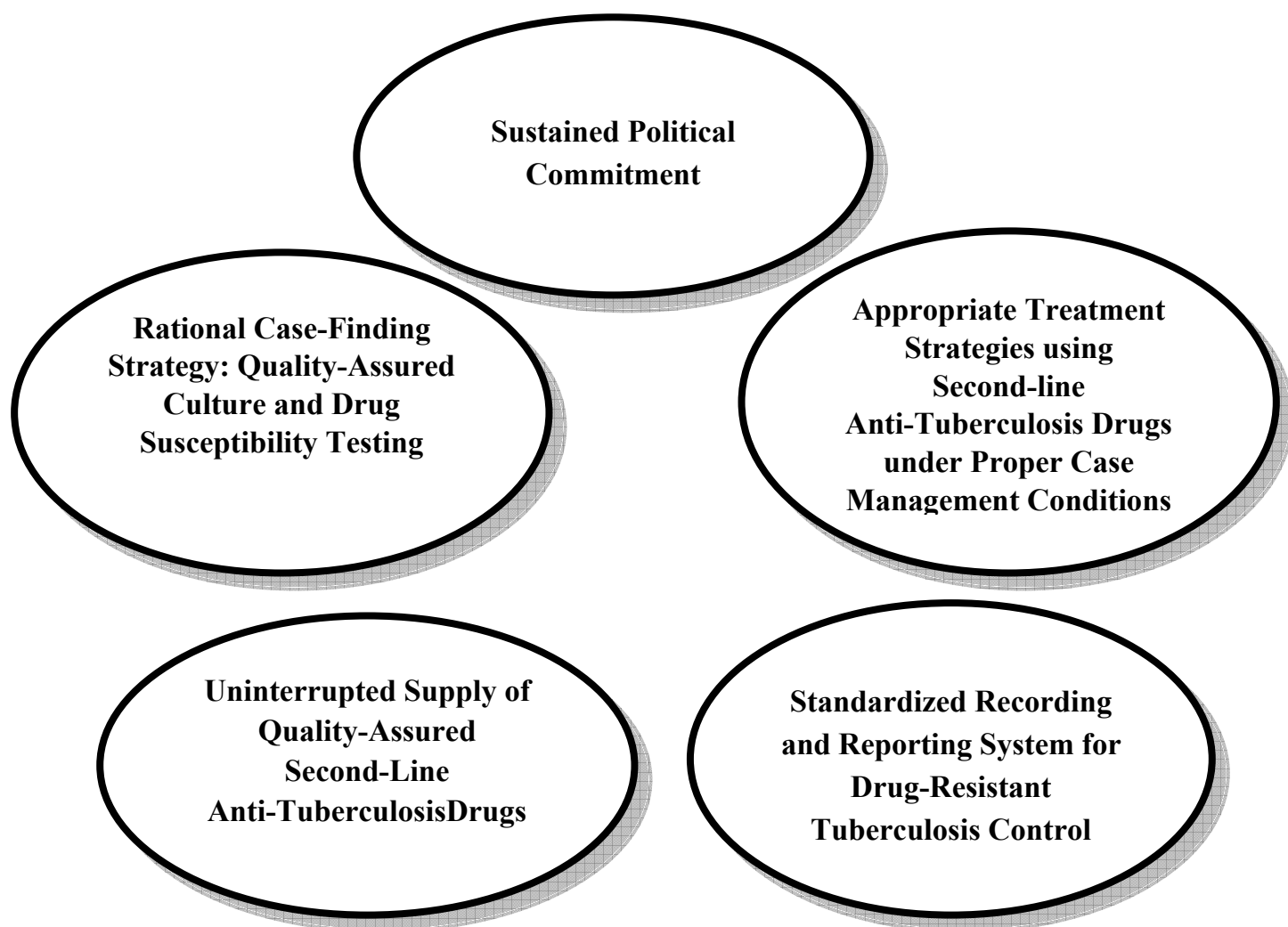
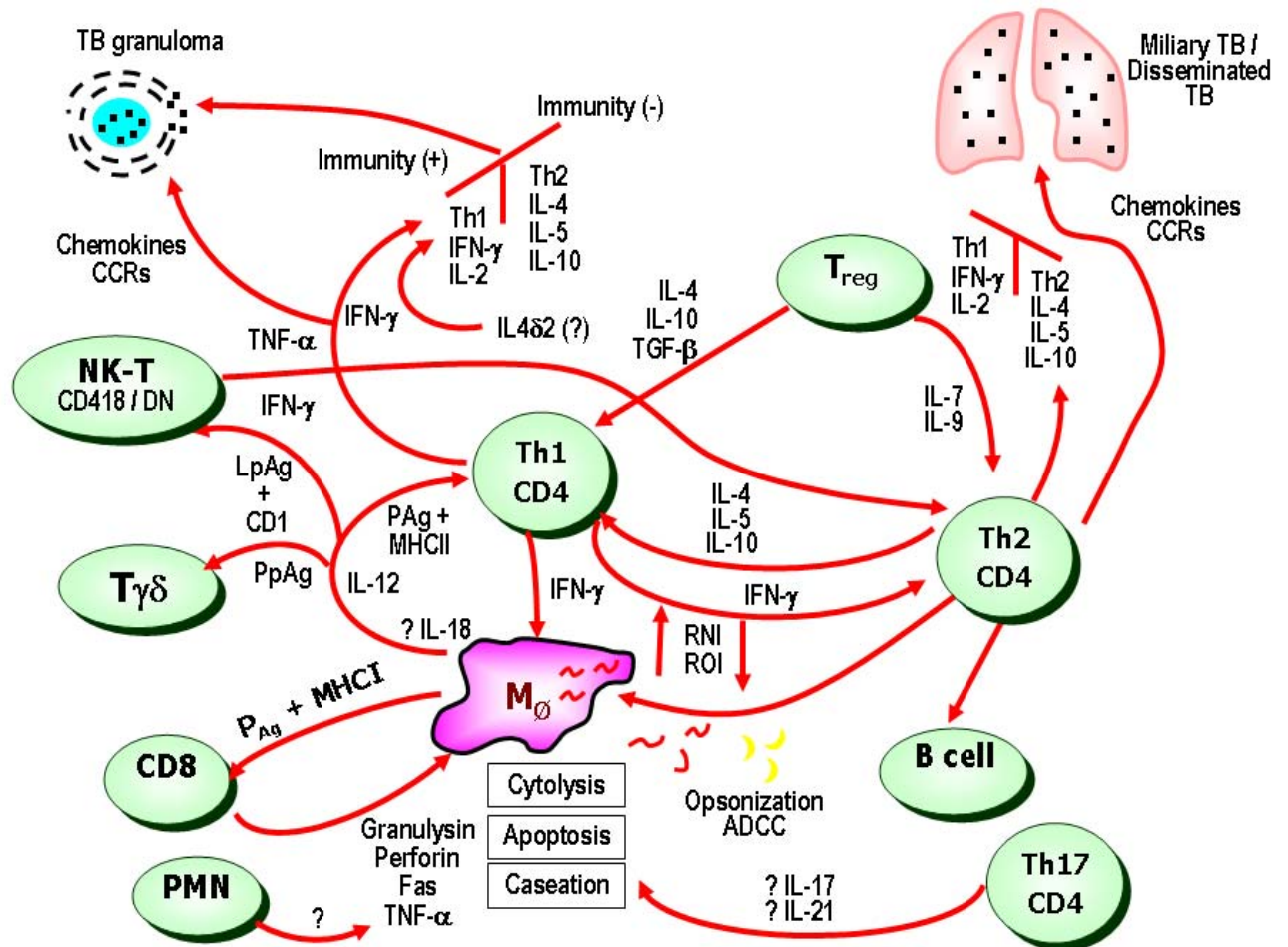


Figure 2 Immunopathogenesis of Tuberculosis



ADCC, antibody-dependent cell-mediated cytotoxicity; CCR, cc chemokine receptor; DN, double negative; Fas, cell receptor inducing apoptosis; IFN-γ, interferon-gamma; IL, interleukin; LpAg, lipopolysaccharide antigen; MHCI, major histocompatibility antigen 1; MHC2, major histocompatibility antigen 2; NK-T, natural killer cell; PAg, peptide antigen; PpAg, phospho-antigen; PMN, polymorphonuclear neutrophil; RNI, reactive nitrogen intermediates; ROI, reactive oxygen intermediates; TGF-β, transforming growth factor-beta; Th, T helper; TNF-α, tumour necrosis factor-alpha; Treg, T regulatory lymphocytes.

Appendix: Grading of Evidences for Recommendations (Scottish Intercollegiate Guidelines Network)

Study Rating	Study Design	Number of Studies	Target Population	Grades of Recommendation
1++	High quality meta-analyses, systematic reviews of RCTs, or RCTs with a very low risk of bias	At least 1 study	Directly Applicable	A
1+	Well-conducted meta-analyses, systematic reviews, or RCTs with a low risk of bias	Studies with overall consistency	Directly Applicable	
1++ / 1+	As above		Extrapolated	
2++	High quality case control/cohort studies or their systemic reviews, with very low risk of confounding/bias and high probability of causal relationship	Studies with overall consistency	Directly Applicable	B
			Extrapolated	
2+	Well-conducted case control/cohort studies with low risk of confounding/bias and a moderate probability of causal relationship	Studies with overall consistency	Directly Applicable	C
			Extrapolated	
3	Non-analytic studies, e.g. case reports, case series			D
4	Expert opinion			

REFERENCES

- [1] World Health Organization. Global tuberculosis control – a short update to the 2009 report. Geneva, Switzerland 2009. WHO/HTM/TB/2009.426.
- [2] World Health Organization. Multidrug and extensively drug-resistant TB (M/XDR-TB): 2010 global report on surveillance and response. Geneva, Switzerland 2010. WHO/HTM/TB/2010.3.
- [3] Scottish Intercollegiate Guidelines Network. SIGN 50: A guideline developer's handbook, Annex B, accessible at <http://www.sign.ac.uk/guidelines/fulltext/50/annexb.html>; last accessed on 26 April 2010.
- [4] Mitchison D. Basic mechanisms of chemotherapy. *Chest* 1979; 76 (suppl): 771 – 81.
- [5] Mitchison DA. The diagnosis and therapy of tuberculosis during the past 100 years. *Am J Respir Crit Care Med* 2005; 171: 699 – 706.
- [6] British Thoracic Society. A controlled trial of 6-months' chemotherapy in pulmonary tuberculosis. Final report: Results during the 36-months after the end of chemotherapy and beyond. *Br J Dis Chest* 1984; 78: 330 – 6.
- [7] Singapore Tuberculosis Service / British Medical Research Council. Five year follow-up of a clinical trial of three 6-month regimens of chemotherapy given intermittently in the continuation phase in the treatment of pulmonary tuberculosis. *Am Rev Respir Dis* 1988; 137: 1147 – 50.
- [8] China tuberculosis control collaboration. Results of directly observed short-course chemotherapy in 112,842 Chinese patients with smear-positive tuberculosis. *Lancet* 1996; 347: 358 – 62.

- [9] Tam CM, Chan SL, Lam CW, et al. Rifapentine and isoniazid in the continuation phase of treating pulmonary tuberculosis: Initial report. *Am J Respir Crit Care Med* 1998; 157: 1726 – 33.
- [10] Snider DE Jr, Graczyk J, Bek E, Rogowski J. Supervised 6-month treatment of newly diagnosed pulmonary tuberculosis using isoniazid, rifampin, and pyrazinamide with and without streptomycin. *Am Rev Respir Dis* 1984; 130: 1091 – 4.
- [11] Third East African / British Medical Research Council Study. Controlled clinical trial of four short-course regimens of chemotherapy for two durations in the treatment of pulmonary tuberculosis. Second Report. *Tubercle* 1980; 61: 59 – 69.
- [12] Hong Kong Chest Service / British Medical Research Council Study. Controlled trial of 6-month and 8-month regimens in the treatment of pulmonary tuberculosis. The results up to 24 months. *Tubercle* 1979; 60: 201 - 10.
- [13] British Thoracic and Tuberculosis Association. Short-course chemotherapy in pulmonary tuberculosis: a controlled trial by the British Thoracic and Tuberculosis Association. *Lancet* 1976; 2: 1102 – 4.
- [14] Slutkin G, Schechter GF, Hopewell PC. The results of 9-month isoniazid-rifampin therapy for pulmonary tuberculosis under program conditions in San Francisco. *Am Rev Respir Dis* 1988; 138: 1622 – 4.
- [15] Combs DL O'Brien RJ Geiter L. USPHS tuberculosis short-course chemotherapy trial 21: effectiveness, toxicity and acceptability — the report of final results. *Ann Intern Med* 1990; 112: 397 – 406.
- [16] World Health Organization. Treatment of tuberculosis. Guidelines: Fourth Edition, Geneva, Switzerland 2010. WHO/HTM/TB/2009.420.

- [17] Quy HT, Cobelens FG, Lan NT, Buu TN, Lambregts CS, Borgdorff MW. Treatment outcomes by drug resistance and HIV status among tuberculosis patients in Ho Chi Minh City, Vietnam. *Int J Tuberc Lung Dis* 2006; 10: 45 – 51.
- [18] Yew WW. Chemotherapy including drug-resistant therapy and future developments in Davies PDO, Barnes PF, Gordon SB (eds). *Clinical tuberculosis* 4th Edition, 2008. Hodder Arnold, U.K., Pp225 – 42.
- [19] Jindani A, Nunn AJ, Enarson DA. Two 8-month regimens of chemotherapy for treatment of newly diagnosed pulmonary tuberculosis: international multicentre randomised trial. *Lancet* 2004; 364: 1244 – 51.
- [20] Menzies D, Benedetti A, Paydar A, et al. Effect of duration and intermittency of rifampin on tuberculosis treatment outcomes: a systematic review and meta-analysis. *PLoS Med* 2009; 6: e1000146.
- [21] Hong Kong Chest Service / British Medical Research Council. Five-year follow-up of a controlled trial of five 6-month regimens of chemotherapy for pulmonary tuberculosis. *Am Rev Respir Dis* 1987; 136: 1339 – 42.
- [22] Hong Kong Chest Service / British Medical Research Council. Controlled trial of 2, 4 and 6 months of pyrazinamide in 6-month three-times-weekly regimens for smear-positive pulmonary tuberculosis, including an assessment of a combined preparation of isoniazid, rifampin and pyrazinamide. Results at 30 months. *Am Rev Respir Dis* 1991; 143: 700 – 6.
- [23] Chang KC, Leung CC, Yew WW, Lau TY, Tam CM. Hepatotoxicity of pyrazinamide: cohort and case-control analyses. *Am J Respir Crit Care Med* 2008; 177: 1391 – 6.
- [24] American Thoracic Society / Centers for Disease Control and Prevention / Infectious Diseases Society of America. Treatment of tuberculosis. *Am J Respir Crit Care Med* 2003; 167: 603 – 62.

- [25] Chang KC, Leung CC, Yew WW, Ho SC, Tam CM. A nested case-control study on treatment-related risk factors for early relapse of tuberculosis. *Am J Respir Crit Care Med* 2004; 170: 1124 – 30.
- [26] Chang KC, Leung CC, Yew WW, Tam CM. Standard anti-tuberculosis treatment and hepatotoxicity: do dosing schedules matter? *Eur Respir J* 2007; 29: 347 – 51.
- [27] Perriens JH, St Louis ME, Mukadi YB, et al. Pulmonary tuberculosis in HIV-infected patients in Zaire. A controlled trial of treatment for either 6 or 12 months. *N Engl J Med*. 1995 23; 332: 779 – 84.
- [28] Nahid P, Gonzalez LC, Rudoy I, et al. Treatment outcomes of patients with HIV and tuberculosis. *Am J Respir Crit Care Med* 2007; 175: 1199 – 206.
- [29] Vernon A, Burman W, Benator D, Khan A, Bozeman L. Acquired rifamycin monoresistance in patients with HIV-related tuberculosis treated with once-weekly rifapentine and isoniazid. Tuberculosis Trials Consortium. *Lancet* 1999; 353: 1843 – 7.
- [30] Burman W, Benator D, Vernon A, Khan A, Jones B, Silva C, Lahart C, Weis S, King B, Mangura B, Weiner M, El-Sadr W. Acquired rifamycin resistance with twice-weekly treatment of HIV-related tuberculosis. *Am J Respir Crit Care Med* 2006; 173: 350 – 6.
- [31] Swaminathan S, Narendran G, Venkatesan P, Iliayas S, Santhanakrishnan R, Menon PA, Padmapriyadarsini C, Ramachandran R, Chinnaiyan P, Suhadev M, Sakthivel R, Narayanan P. Efficacy of a 6 vs. a 9-month Intermittent Treatment Regimen in HIV-infected TB Patients: A Randomized Clinical Trial. *Am J Respir Crit Care Med* 2010; 181: 743 – 51.
- [32] Hong Kong Chest Service / Tuberculosis Research Centre, Madras / British Medical Research Council. A controlled trial of 2-month, 3-month and 12-month regimens of chemotherapy for sputum-smear-negative pulmonary tuberculosis: results at 60 months. *Am Rev Respir Dis* 1984; 130: 23 – 8.

- [33] Hong Kong Chest Service / Tuberculosis Research Centre, Madras / British Medical Research Council. A controlled trial of 3-month, 4-month and 6-month regimens of chemotherapy for sputum-smear-negative pulmonary tuberculosis. Results at 5 years. *Am Rev Respir Dis* 1989; 139: 871 – 6.
- [34] Nunn AJ, Phillips PP, Mitchison DA. Timing of relapse in short-course chemotherapy trials for tuberculosis. *Int J Tuberc Lung Dis* 2010; 14: 241 – 2.
- [35] Kochi A. Is DOTS the health breakthrough of the 1990s? *World Health Forum* 1997; 18: 225 – 47.
- [36] Yew WW. Directly observed therapy, short-course: the best way to prevent multidrug-resistant tuberculosis. *Chemotherapy* 1999; 45 (suppl 2): S26 – 33.
- [37] Sumartojo E. When tuberculosis treatment fails. A social behavioral account of patient adherence. *Am Rev Respir Dis* 1993; 147: 1311 – 20.
- [38] Volmink J, Garner P. Directly observed therapy for treating tuberculosis. *Cochrane Database Syst Rev* 2007; 4: CD003343.
- [39] Rusen ID. Cochrane systematic review of directly observed therapy for treating tuberculosis: good analysis of the wrong outcome. *Int J Tuberc Lung Dis* 2007; 11: 120 – 1.
- [40] Thiam S, LeFevre AM, Hane F, et al. Effectiveness of a strategy to improve adherence to tuberculosis treatment in a resource-poor setting: a cluster randomized controlled trial. *JAMA* 2007; 297: 380 – 6.
- [41] Mishra P, Hansen EH, Sabroe S, Kafle KK. Adherence is associated with the quality of professional-patient interaction in Directly Observed Treatment Short-course DOTS. *Patient Educ Couns* 2006; 63: 29 – 37.

- [42] Newell JN, Baral SC, Pande SB, Bam DS, Malla P. Family-member DOTS and community DOTS for tuberculosis control in Nepal: cluster-randomised controlled trial. *Lancet* 2006; 367: 903 – 9.
- [43] Sbarbaro J, Blomberg B, Chaulet P. Fixed-dose combination formulations for tuberculosis treatment. *Int J Tuberc Lung Dis* 1999; 3: S286 – 8.
- [44] Moulding T, Dutt AK, Reichman LB. Fixed-dose combinations of antituberculous medications to prevent drug resistance. *Ann Intern Med* 1995; 122: 951 – 4.
- [45] Moulding TS, Le HQ, Rikleen D, Davidson P. Preventing drug-resistant tuberculosis with a fixed dose combination of isoniazid and rifampicin. *Int J Tuberc Lung Dis* 2004; 8: 743 – 8.
- [46] WHO model list of essential drugs, available at www.who.int/medicines/publications/essentialmedicines/en; last accessed on 26 April 2010.
- [47] A joint statement of the International Union Against Tuberculosis and Lung Disease / and the Tuberculosis Programme of the World Health Organization. The promise and reality of fixed-dose combinations with rifampicin. *Tubercle Lung Dis* 1994; 75: 180 – 1.
- [48] ERS, WHO, IUATLD (Europe Region) Task Force. Tuberculosis management in Europe: recommendations of a task force of the European Respiratory Society, the World Health Organization and the International Union against Tuberculosis and Lung Disease (Europe Region). *Eur Respir J* 1999; 14: 978 – 92.
- [49] Bartacek A, Schutt D, Panosch B, Borek M. Rimstar 4-FDC Study Group. Comparison of a four-drug fixed-dose combination regimen with a single tablet regimen in smear-positive pulmonary tuberculosis. *Int J Tuberc Lung Dis* 2009; 13: 760 – 6.

- [50] Teo SK. Assessment of a combined preparation of isoniazid, rifampicin and pyrazinamide (Rifater ®) in the initial phase of chemotherapy in three 6-month regimens for smear positive pulmonary tuberculosis: a five-year follow-up report. *Int J Tuberc Lung Dis* 1999; 3: 126 – 32.
- [51] Mitchison DA. How drug resistance emerges as a result of poor compliance during short course chemotherapy for tuberculosis. *Int J Tuberc Lung Dis* 1998; 2: 10 – 5.
- [52] Sirgel FA, Fourie PB, Donald PR, et al. The early bactericidal activities of rifampin and rifapentine in pulmonary tuberculosis. *Am J Respir Crit Care Med* 2005; 172: 128 – 35.
- [53] Diacon AH, Patientia RF, Venter A, et al. Early bactericidal activity of high-dose rifampin in patients with pulmonary tuberculosis evidenced by positive sputum smears. *Antimicrob Agents Chemother* 2007; 51: 2994 – 6.
- [54] Gumbo T, Louie A, Deziel MR, et al. Concentration-dependent *Mycobacterium tuberculosis* killing and prevention of resistance by rifampin. *Antimicrob Agents Chemother* 2007; 51: 3781 – 8.
- [55] Mitnick CD, McGee B, Peloquin CA. Tuberculosis pharmacotherapy: strategies to optimize patient care. *Expert Opin Pharmacother* 2009; 10: 381 – 401.
- [56] Saukkonen JJ, Cohn DL, Jasmer RM, et al. An official ATS statement: hepatotoxicity of antituberculosis therapy. *Am J Respir Crit Care Med* 2006; 174: 935 – 52.
- [57] Steele MA, Burk RF, DesPrez RM. Toxic hepatitis with isoniazid and rifampin. A meta-analysis. *Chest* 1991; 99: 465 – 71.
- [58] Ruslami R, Nijland HM, Alisjahbana B, Parwati I, van Crevel R, Aarnoutse RE. Pharmacokinetics and tolerability of a higher rifampin dose versus the standard dose in pulmonary tuberculosis patients. *Antimicrob Agents Chemother* 2007; 51: 2546 – 51.

- [59] Long MW, Snider DE Jr, Farer LS. U.S. Public Health Service Cooperative Trial of three rifampin-isoniazid regimens in treatment of pulmonary tuberculosis. *Am Rev Respir Dis* 1979; 119: 879 – 94.
- [60] Peloquin C. What is the “right” dose of rifampin? *Int J Tuberc Lung Dis* 2003; 7: 3 – 5.
- [61] Pharmacokinetics and pharmacodynamics of high versus standard dose rifampicin in patients with pulmonary tuberculosis (High RIF) registered at <http://clinicaltrials.gov/ct2/show/NCT00760149> last accessed on 26 April 2010.
- [62] Tam CM, Chan SL, Kam KM, Goodall RL, Mitchison DA. Rifapentine and isoniazid in the continuation phase of a 6-month regimen. Final report at 5 years: prognostic value of various measures. *Int J Tuberc Lung Dis* 2002; 6: 3 – 10.
- [63] Benator D, Bhattacharya M, Bozeman L, et al. Tuberculosis Trials Consortium Rifapentine and isoniazid once a week versus rifampicin and isoniazid twice a week for treatment of drug-susceptible pulmonary tuberculosis in HIV-negative patients: a randomized clinical trial. *Lancet* 2002; 360: 528 – 34.
- [64] Langdon G, Wilkins JJ, Smith PJ, McIlleron H. Consecutive-dose pharmacokinetics of rifapentine in patients diagnosed with pulmonary tuberculosis. *Int J Tuberc Lung Dis*. 2004;8:862-7.
- [65] Weiner M, Bock N, Peloquin CA, et al. Tuberculosis Trials Consortium. Pharmacokinetics of rifapentine at 600, 900, and 1200 mg during once-weekly tuberculosis therapy. *Am J Respir Crit Care Med* 2004; 169: 1191 – 7.
- [66] Rosenthal IM, Williams K, Tyagi S, et al. Potent twice-weekly rifapentine-containing regimens in murine tuberculosis. *Am J Respir Crit Care Med* 2006; 174: 94 – 101.
- [67] Rosenthal IM, Zhang M, Williams KN. Daily dosing of rifapentine cures tuberculosis in three months or less in the murine model. *PLoS Med* 2007; 4: e344.

- [68] Dooley K, Flexner C, Hackman J, et al. Repeated administration of high-dose intermittent rifapentine reduces rifapentine and moxifloxacin plasma concentrations. *Antimicrob Agents Chemother* 2008; 52: 4037 – 42.
- [69] Bock NN, Sterling TR, Hamilton CD, et al. Tuberculosis Trials Consortium, Centers for Disease Control and Prevention, Atlanta, Georgia. A prospective, randomized, double-blind study of the tolerability of rifapentine 600, 900, and 1200 mg plus isoniazid in the continuation phase of tuberculosis treatment. *Am J Respir Crit Care Med* 2002; 165: 1526 – 30.
- [70] TBTC Study 29: Rifapentine during intensive phase tuberculosis (TB) treatment registered at <http://clinicaltrials.gov/ct2/show/NCT00694629>; last accessed on 26 April 2010.
- [71] Caminero JA. Multidrug-resistant tuberculosis: epidemiology, risk factors and case finding. *Int J Tuberc Lung Dis* 2010; 14: 382 – 90.
- [72] Yoshiyama T, Yanai H, Rhiengtong D, et al. Development of acquired drug resistance in recurrent tuberculosis patients with various previous treatment outcomes. *Int J Tuberc Lung Dis* 2004; 8: 31 – 8.
- [73] Davidson PT. Drug resistance and the selection of therapy for tuberculosis. *Am Rev Respir Dis* 1987; 136: 255 – 7.
- [74] Joint Tuberculosis Committee of the British Thoracic Society. Chemotherapy and management of tuberculosis in the United Kingdom: recommendations 1998. *Thorax* 1998; 53: 536 – 48.
- [75] Mitchison DA, Nunn AJ. Influence of initial drug resistance on the response to short-course chemotherapy of pulmonary tuberculosis. *Am Rev Respir Dis* 1986; 133: 423 – 30.

- [76] Barnes PF, Bloch AB, Davidson PT, Snider DE Jr. Tuberculosis in patients with human immunodeficiency virus infection. *N Engl J Med* 1991; 324: 1644 – 50.
- [77] Li J, Munsiff SS, Driver CR, Sackoff J. Relapse and acquired rifampin resistance in HIV-infected patients with tuberculosis treated with rifampin- or rifabutin-based regimens in New York City 1997 – 2000. *Clin Infect Dis* 2005; 41: 83 – 91.
- [78] Traore H, Fissette K, Bastian I, Devleeschouwer M, Portaels F. Detection of rifampicin resistance in *Mycobacterium tuberculosis* isolates from diverse countries by a commercial line probe assay as an initial indicator of multidrug resistance. *Int J Tuberc Lung Dis* 2000; 4: 481 – 4.
- [79] Yam WC, Tam CM, Leung CC, et al. Direct detection of rifampin-resistant *Mycobacterium tuberculosis* in respiratory specimens by PCR-DNA sequencing. *J Clin Microbiol* 2004; 42: 4438 – 43.
- [80] Espinal MA, Kim SJ, Suarez PG, et al. Standard short-course chemotherapy for drug-resistant tuberculosis: treatment outcomes in 6 countries. *JAMA* 2000; 283: 2537 – 45.
- [81] Becerra MC, Freeman J, Bayona J, et al. Using treatment failure under effective directly observed short-course chemotherapy programs to identify patients with multidrug-resistant tuberculosis. *Int J Tuberc Lung Dis* 2000; 4: 108 – 14.
- [82] Mak A, Thomas A, Del Granado M, Zaleskis R, Mouzafarova N, Menzies D. Influence of multidrug resistance on tuberculosis treatment outcomes with standardized regimens. *Am J Respir Crit Care Med* 2008; 178: 306 – 12.
- [83] World Health Organization. Guidelines for the programmatic management of drug-resistant tuberculosis. Geneva, Switzerland. 2006. WHO/HTM/TB/2006.361.

- [84] Caminero J A. Likelihood of generating MDR-TB and XDR-TB under adequate National Tuberculosis Control Programme implementation. *Int J Tuberc Lung Dis* 2008; 12: 869 – 77.
- [85] Louw GE, Warren RM, Donald PR, et al. Frequency and implications of pyrazinamide resistance in managing previously treated tuberculosis patients. *Int J Tuberc Lung Dis* 2006; 10: 802 – 7.
- [86] Hanif M, Malik S, Dhingra VK. Acquired drug resistance pattern in tuberculosis cases at the State Tuberculosis Centre, Delhi, India. *Int J Tuberc Lung Dis* 2009; 13: 74 – 8.
- [87] Migliori GB, Besozzi G, Girardi E, et al. Clinical and operational value of the extensively drug-resistant tuberculosis definition. *Eur Respir J* 2007; 30: 623 – 6.
- [88] Yew WW, Chan CK, Chau CH, et al. Outcomes of patients with multidrug-resistant pulmonary tuberculosis treated with ofloxacin / levofloxacin-containing regimens. *Chest* 2000; 117: 744 – 51.
- [89] Chan ED, Laurel V, Strand MJ, et al. Treatment and outcome analysis of 205 patients with multidrug-resistant tuberculosis. *Am J Respir Crit Care Med* 2004; 169: 1103 – 9.
- [90] Chiang CY, Enarson DA, Yu MC, et al. Outcome of pulmonary multidrug-resistant tuberculosis: a 6-yr follow-up study. *Eur Respir J* 2006; 28: 980 – 5.
- [91] Park SK, Lee WC, Lee DH, Mitnick CD, Han L, Seung KJ. Self-administered, standardized regimens for multidrug-resistant tuberculosis in South Korea. *Int J Tuberc Lung Dis* 2004; 8: 361 – 8.
- [92] Leimane V, Riekstina V, Holtz TH, et al. Clinical outcome of individualised treatment of multidrug-resistant tuberculosis in Latvia: a retrospective cohort study. *Lancet* 2005; 365: 318 – 26.

- [93] Huang TS, Kunin CM, Shin-Jung Lee S, Chen YS, Tu HZ, Liu YC. Trends in fluoroquinolone resistance of *Mycobacterium tuberculosis* complex in a Taiwanese medical centre: 1995 – 2003. *J Antimicrob Chemother* 2005; 56: 1058 – 62.
- [94] Park IN, Hong SB, Oh YM, et al. Impact of short-term exposure to fluoroquinolones on ofloxacin resistance in HIV-negative patients with tuberculosis. *Int J Tuberc Lung Dis* 2007; 11: 319 – 24.
- [95] Devasia RA, Blackman A, Gebretsadik T, et al. Fluoroquinolones resistance in *Mycobacterium tuberculosis*: the effect of duration and timing of fluoroquinolone exposure. *Am J Respir Crit Care Med* 2009; 180: 365 – 70.
- [96] Migliori GB, Lange C, Centis R, et al. Resistance to second-line injectables and treatment outcomes in multidrug-resistant and extensively drug-resistant tuberculosis cases. *Eur Respir J* 2008; 31: 1155 – 9.
- [97] World Health Organization. Global Tuberculosis Control – surveillance, planning, financing. Geneva, Switzerland: WHO 2008. WHO/HTM/TB/2008.393.
- [98] Dewan P, Sosnovskaja A, Thomsen V, et al. High prevalence of drug-resistant tuberculosis, Republic of Lithuania, 2002. *Int J Tuberc Lung Dis* 2005; 9: 170 – 4.
- [99] Timperi R, Han LL, Sloutsky A, et al. Drug resistance profiles of *Mycobacterium tuberculosis* isolates: five years' experience and insight into treatment strategies for MDR-TB in Lima, Peru. *Int J Tuberc Lung Dis* 2005; 9: 175 – 80.
- [100] World Health Organization. Guidelines for Drug Susceptibility Testing for Second-line Anti-tuberculosis Drugs for DOTS-plus. Geneva, Switzerland. 2001. WHO/CDS/TB/2001.288.
- [101] Drobniewski F, Rusch-Gerdes S, Hoffner S. Antimicrobial susceptibility testing of *Mycobacterium tuberculosis* (EUCAST document E.DEF.8.1) – report of the Subcommittee on Antimicrobial Susceptibility Testing of *Mycobacterium tuberculosis*

- of the European Committee for Antimicrobial Susceptibility Testing (EUCAST) of the European Society of Clinical Microbiology and Infectious Diseases (ESCMID). *Clin Microbiol Infect* 2007; 13: 1144 – 56.
- [102] Kim SJ. Drug-susceptibility testing in tuberculosis: methods and reliability of results. *Eur Respir J* 2005; 25: 564 – 9.
- [103] Kam KM, Sloutsky A, Yip CW, et al. Determination of critical concentrations of second-line anti-tuberculosis drugs with clinical and microbiological relevance. *Int J Tuberc Lung Dis* 2010; 14: 282 – 8.
- [104] Kang YA, Choi YJ, Cho YJ, et al. Cost of treatment for multidrug-resistant tuberculosis in South Korea. *Respirology* 2006; 11: 793 – 8.
- [105] Sterling TR, Lehmann HP, Frieden TR. Impact of DOTS compared with DOTS-Plus on multidrug resistant tuberculosis and tuberculosis deaths: decision analysis. *BMJ* 2003; 326: 574.
- [106] Tupasi TE, Gupta R, Quelapio MI, et al. Feasibility and cost-effectiveness of treating multidrug-resistant tuberculosis: a cohort study in the Philippines. *PLoS Med.* 2006; 3: e352.
- [107] Nathanson E, Lambregts-van Weezenbeek C, Rich ML, et al. Multidrug-resistant tuberculosis management in resource-limited settings. *Emerg Infect Dis* 2006; 12: 1389 – 97.
- [108] Bastian I, Rigouts L, Van Deun A, Portaels F. Directly observed treatment, short-course strategy and multidrug-resistant tuberculosis: are any modifications required? *Bull World Health Organ* 2000; 78: 238 – 51.
- [109] Gupta R, Cegielski JP, Espinal MA, et al. Increasing transparency in partnerships for health – introducing the Green Light Committee. *Trop Med Int Health* 2002; 7: 970 – 6.

- [110] Caminero JA. Treatment of multidrug-resistant tuberculosis: evidence and controversies. *Int J Tuberc Lung Dis* 2006; 10: 829 – 37.
- [111] Mitnick CD, Castro KG, Harrington M, Sacks LV, Burman W. Randomized trials to optimize treatment of multidrug-resistant tuberculosis. *PLoS Med* 2007; 4: e282.
- [112] World Health Organization. Guidelines for the Programmatic Management of Drug-Resistant Tuberculosis – Emergency Update 2008. Geneva, Switzerland. 2008. WHO/HTM/TB/2008.402.
- [113] Katiyar SK, Bihari S, Prakash S, Mamtani M, Kulkarni H. A randomised controlled trial of high-dose isoniazid adjuvant therapy for multidrug-resistant tuberculosis. *Int J Tuberc Lung Dis* 2008; 12: 139 – 45.
- [114] Migliori GB, Lange C, Girardi E, et al. Fluoroquinolones: are they essential to treat multidrug-resistant tuberculosis? *Eur Respir J* 2008; 31: 904 – 5.
- [115] Devasia RA, Blackman A, May C, et al. Fluoroquinolone resistance in *Mycobacterium tuberculosis*: an assessment of MGIT 960, MODS, and nitrate reductase assay and fluoroquinolone cross-resistance. *J Antimicrob Chemother* 2009; 63: 1173 – 8.
- [116] Moadebi S, Harder CK, Fitzgerald MJ, Elwood KR, Marra F. Fluoroquinolones for the treatment of pulmonary tuberculosis. *Drugs* 2007; 67: 2077 – 99.
- [117] Yew WW, Chan CK, Leung CC, et al. Comparative roles of levofloxacin and ofloxacin in the treatment of multidrug-resistant tuberculosis: preliminary results of a retrospective study from Hong Kong. *Chest* 2003; 124: 1476 – 81.
- [118] Gumbo T, Louie A, Deziel MR, Parsons LM, Salfinger M, Drusano GL. Selection of a moxifloxacin dose that suppresses drug resistance in *Mycobacterium tuberculosis*, by use of an *in vitro* pharmacodynamic infection model and mathematical modeling. *J Infect Dis* 2004; 190: 1642 – 51.

- [119] Allen BW, Mitchison DA, Chan YC, Yew WW, Allan WG, Girling DJ. Amikacin in the treatment of pulmonary tuberculosis. *Tubercle* 1983; 64: 111 – 8.
- [120] McClatchy JK, Kanes W, Davidson PT, Moulding TS. Cross-resistance in *M. tuberculosis* to kanamycin, capreomycin and viomycin. *Tubercle* 1977; 58: 29 – 34.
- [121] Jeon DS, Kim DH, Kang HS, et al. Survival and predictors of outcomes in non-HIV-infected patients with extensively drug-resistant tuberculosis. *Int J Tuberc Lung Dis* 2009; 13: 594 – 600.
- [122] Zurenko GE, Yagi BH, Schaadt RD, et al. *In vitro* activities of U-100592 and U-100766, novel oxazolidinone antibacterial agents. *Antimicrob Agents Chemother* 1996; 40: 839 – 45.
- [123] von der Lippe B, Sandven P, Brubakk O. Efficacy and safety of linezolid in multidrug-resistant tuberculosis (MDR-TB) – a report of ten cases. *J Infect* 2006; 52: 92 – 6.
- [124] Park IN, Hong SB, Oh YM, et al. Efficacy and tolerability of daily-half dose linezolid in patients with intractable multidrug-resistant tuberculosis. *J Antimicrob Chemother* 2006; 58: 701 – 4.
- [125] Boutoille D, Grossi O, Depatureaux A, Tattevin P. Fatal lactic acidosis after prolonged linezolid exposure for treatment of multidrug-resistant tuberculosis. *Eur J Intern Med* 2009; 20: e134 – 5.
- [126] Migliori GB, Eker B, Richardson MD, et al. TBNET Study Group. A retrospective TBNET assessment of linezolid safety, tolerability and efficacy in multidrug-resistant tuberculosis. *Eur Respir J* 2009; 34: 387 – 93.
- [127] TBTC S30: Safety and tolerability of low dose linezolid in MDR TB (LiMiT), registered at <http://clinicaltrials.gov/ct2/show/NCT00664313>; last accessed on 26 April 2010.

- [128] Koh WJ, Kwon OJ, Gwak H, et al. Daily 300 mg dose of linezolid for the treatment of intractable multidrug-resistant and extensively drug-resistant tuberculosis. *J Antimicrob Chemother* 2009; 64: 388 – 91.
- [129] Yew WW, Chang KC, Chau CH. Comment on : Daily 300 mg dose of linezolid for the treatment of intractable multidrug-resistant and extensively drug-resistant tuberculosis. *J Antimicrob Chemother* 2009; 64: 1119.
- [130] Williams KN, Stover CK, Zhu T, et al. Promising anti-tuberculosis activity of the oxazolidinone PNU-100480 relative to linezolid in the murine model. *Antimicrob Agents Chemother* 2009; 53: 1314 – 9.
- [131] Study to evaluate safety, tolerability and pharmacokinetics (PK) (Including food effect) of oral doses of AZD5847 in healthy volunteers; registered at <http://clinicaltrials.gov/ct2/show/NCT01037725>; last accessed on 26 April 2010.
- [132] Donald PR, Sirgel FA, Venter A, et al. Early bactericidal activity of amoxicillin in combination with clavulanic acid in patients with sputum smear-positive pulmonary tuberculosis. *Scand J Infect Dis* 2001; 33: 466 – 9.
- [133] Abate G, Miorner H. Susceptibility of multidrug-resistant strains of *Mycobacterium tuberculosis* to amoxicillin in combination with clavulanic acid and ethambutol. *J Antimicrob Chemother* 1998; 42: 735 – 40.
- [134] Dincer I, Ergin A, Kocagoz T. The *in vitro* efficacy of β -lactam and β -lactamase inhibitors against multidrug-resistant clinical strains of *Mycobacterium tuberculosis*. *Int J Antimicrob Agents* 2004; 23: 408 – 11.
- [135] Nadler JP, Berger J, Nord JA, Cofsky R, Saxena M. Amoxicillin-clavulanic acid for treating drug-resistant *Mycobacterium tuberculosis*. *Chest* 1991; 99: 1025 – 6.

- [136] Yew WW, Wong CF, Lee J, Wong PC, Chau CH. Do β -lactam- β -lactamase inhibitor combinations have a place in the treatment of multidrug-resistant pulmonary tuberculosis? *Tuberc Lung Dis* 1995; 76: 90 – 2.
- [137] Chambers HF, Moreau D, Yajko D, et al. Can penicillins and other beta-lactam antibiotics be used to treat tuberculosis? *Antimicrob Agents Chemother* 1995; 39: 2620 – 4.
- [138] Chambers HF, Turner J, Schechter GF, Kawamura M, Hopewell PC. Imipenem for treatment of tuberculosis in mice and humans. *Antimicrob Agents Chemother* 2005; 49: 2816 – 21.
- [139] Hugonnet JE, Tremblay LW, Boshoff HI, Barry CE 3rd, Blanchard JS. Meropenem-clavulanate is effective against extensively drug-resistant *Mycobacterium tuberculosis*. *Science* 2009; 323: 1215 – 8.
- [140] Holzgrabe U. Meropenem-clavulanate: a new strategy for the treatment of tuberculosis? *Chem Med Chem* 2009; 4: 1051 – 3.
- [141] Jagannath C, Reddy MV, Kailasam S, O'Sullivan JF, Gangadharam PR. Chemotherapeutic activity of clofazimine and its analogues against *Mycobacterium tuberculosis*. *In vitro*, intracellular and *in vivo* studies. *Am J Respir Crit Care Med* 1995; 151: 1083 – 6.
- [142] Matlola NM, Steel HC, Anderson R. Antimycobacterial action of B4128, a novel tetramethylpiperidyl-substituted phenazine. *J Antimicrob Chemother* 2001; 47: 199 – 202.
- [143] Janulionis E, Sofer C, Song HY, Wallis RS. Lack of activity of orally administered clofazimine against intracellular *Mycobacterium tuberculosis* in whole-blood culture. *Antimicrob Agents Chemother* 2004; 48: 3133 – 5.

- [144] Uskudar O, Koksul D, Koksul AS. Partial intestinal obstruction due to clofazimine in a patient with multidrug-resistant tuberculosis. *Int J Tuberc Lung Dis* 2005; 9: 703 – 4.
- [145] Richter E, Rusch-Gerdes S, Hillemann D. First linezolid-resistant clinical isolates of *Mycobacterium tuberculosis*. *Antimicrob Agents Chemother* 2007; 51: 1534 – 6.
- [146] Huang TS, Liu YC, Sy CL, Chen YS, Tu HZ, Chen BC. *In vitro* activities of linezolid against clinical isolates of *Mycobacterium tuberculosis* complex isolated in Taiwan over 10 years. *Antimicrob Agents Chemother* 2008; 52: 2226 – 7.
- [147] Yew WW, Leung CC. Management of multidrug-resistant tuberculosis. Update 2007. *Respirology* 2008; 13: 21 – 46.
- [148] Johnson JL, Hadad DJ, Boon WH, et al. Early and extended early bactericidal activity of levofloxacin, gatifloxacin and moxifloxacin in pulmonary tuberculosis. *Int J Tuberc Lung Dis* 2006; 10: 605 – 12.
- [149] Perez-Guzman C, Vargas MH, Martinez-Rossier LA, Torres-Cruz A, Villarreal-Velarde H. Results of a 12-month regimen for drug-resistant pulmonary tuberculosis. *Int J Tuberc Lung Dis* 2002; 6: 1102 – 9.
- [150] Furin JJ, Mitnick CD, Shin SS, et al. Occurrence of serious adverse effects in patients receiving community-based therapy for multidrug-resistant tuberculosis. *Int J Tuberc Lung Dis* 2001; 5: 648 – 55.
- [151] Torun T, Gungor G, Ozmen I, et al. Side effects associated with the treatment of multidrug-resistant tuberculosis. *Int J Tuberc Lung Dis* 2005; 9: 1373 – 7.
- [152] Coyne KM, Pozniak AL, Lamorde M, Boffito M. Pharmacology of second-line antituberculosis drugs and potential for interactions with antiretroviral agents *AIDS* 2009; 23; 437-446.

- [153] de Jager P, van Altena R. Hearing loss and nephrotoxicity in long-term aminoglycoside treatment in patients with tuberculosis. *Int J Tuberc Lung Dis* 2002; 6: 622 – 7.
- [154] Goble M, Iseman MD, Madsen LA, Waite D, Ackerson L, Horsburgh CR Jr. Treatment of 171 patients with pulmonary tuberculosis resistant to isoniazid and rifampin. *N Engl J Med* 1993; 328: 527 – 32.
- [155] Telzak EE, Sepkowitz K, Alpert P, et al. Multidrug-resistant tuberculosis in patients without HIV infection. *N Engl J Med* 1995; 333: 907 – 11.
- [156] Park MM, Davis AL, Schluger NW, Cohen H, Rom WN. Outcome of MDR-TB patients, 1983 – 1993. Prolonged survival with appropriate therapy. *Am J Respir Crit Care Med* 1996; 153: 317 – 24.
- [157] Park SK, Kim CT, Song SD. Outcome of chemotherapy in 107 patients with pulmonary tuberculosis resistant to isoniazid and rifampin. *Int J Tuberc Lung Dis* 1998; 2: 877 – 84.
- [158] Tahaoglu K, Torun T, Sevim T, et al. The treatment of multidrug-resistant tuberculosis in Turkey. *N Engl J Med* 2001; 345: 170 – 4.
- [159] Drobniewski F, Eltringham I, Graham C, Magee JG, Smith EG, Watt B. A national study of clinical and laboratory factors affecting the survival of patients with multiple drug resistant tuberculosis in the UK. *Thorax* 2002; 57: 810 – 6.
- [160] Mitnick C, Bayona J, Palacios E, et al. Community-based therapy for multidrug-resistant tuberculosis in Lima, Peru. *N Engl J Med* 2003; 348: 119 – 28.
- [161] Pearson ML, Jereb JA, Frieden TR, et al. Nosocomial transmission of multidrug-resistant *Mycobacterium tuberculosis*: a risk to patients and healthcare workers. *Ann Intern Med* 1992; 117: 191 – 6.

- [162] Edlin BR, Tokars JI, Grieco MH, et al. An outbreak of multidrug-resistant tuberculosis among hospitalized patients with the acquired immunodeficiency syndrome. *N Engl J Med* 1992; 326: 1514 – 21.
- [163] Franke MF, Appleton SC, Bayona J, et al. Risk factors and mortality associated with default from multidrug-resistant tuberculosis treatment. *Clin Infect Dis* 2008; 46: 1844 – 51.
- [164] Kim HR, Hwang SS, Kim HJ, et al. Impact of extensive drug resistance on treatment outcomes in non-HIV-infected patients with multidrug-resistant tuberculosis. *Clin Infect Dis* 2007; 45: 1290 – 5.
- [165] Cox HS, Kalon S, Alamuratova S, et al. Multidrug-resistant tuberculosis treatment outcomes in Karakalpakstan, Uzbekistan: treatment complexity and XDR-TB among treatment failures. *PLoS One* 2007; 2: e1126.
- [166] Gandhi NR, Shah NS, Andrews JR, et al. HIV co-infection in multidrug- and extensively drug-resistant tuberculosis results in high early mortality. *Am J Respir Crit Care Med* 2010; 181: 80 – 6.
- [167] Kliiman K, Altraja A. Predictors of poor treatment outcome in multidrug- and extensively drug-resistant pulmonary TB. *Eur Respir J* 2009; 33: 1085 – 94.
- [168] Orenstein EW, Basu S, Shah NS, et al. Treatment outcomes among patients with multidrug-resistant tuberculosis: systematic review and meta-analysis. *Lancet Infect Dis* 2009; 9: 153 – 61.
- [169] Mitnick CD, Shin SS, Seung KJ, et al. Comprehensive treatment of extensively drug-resistant tuberculosis. *N Engl J Med* 2008; 359: 563 – 74.
- [170] Sotgiu G, Ferrara G, Matteelli MD, et al. Epidemiology and clinical management of XDR-TB: a systematic review by TBNET. *Eur Respir J* 2009; 33: 871 – 81.

- [171] Spigelman MK. New tuberculosis therapeutics: a growing pipeline. *J Infect Dis* 2007; 196: Suppl 1: S28 – 34.
- [172] Nuermberger EL, Yoshimatsu T, Tyagi S, et al. Moxifloxacin-containing regimen greatly reduces time to culture conversion in murine tuberculosis. *Am J Respir Crit Care Med* 2004; 169: 421 – 6.
- [173] Nuermberger EL, Yoshimatsu T, Tyagi S, et al. Moxifloxacin-containing regimens of reduced duration produce a stable cure in murine tuberculosis. *Am J Respir Crit Care Med* 2004; 170: 1131 – 4.
- [174] Burman WJ, Goldberg S, Johnson JL, et al. Moxifloxacin versus ethambutol in the first 2 months of treatment for pulmonary tuberculosis. *Am J Respir Crit Care Med* 2006; 174: 331 – 8.
- [175] Conde MB, Efron A, Loredó C, et al. Moxifloxacin versus ethambutol in the initial treatment of tuberculosis: a double-blind, randomised, controlled phase II trial. *Lancet* 2009; 373: 1183 – 9.
- [176] Rustomjee R, Lienhardt C, Kanyok T, et al. A Phase II study of the sterilizing activities of ofloxacin, gatifloxacin and moxifloxacin in pulmonary tuberculosis. *Int J Tuberc Lung Dis* 2008; 12: 128 – 38.
- [177] Dorman SE, Johnson JL, Goldsberg S, et al. Substitution of moxifloxacin for isoniazid during intensive phase treatment of pulmonary tuberculosis. *Am J Respir Crit Care Med* 2009; 180: 273 – 80.
- [178] Wang JY, Wang JT, Tsai TH, et al. Adding moxifloxacin is associated with a shorter time to culture conversion in pulmonary tuberculosis. *Int J Tuberc Lung Dis*. 2010; 14: 65 – 71.

- [179] Controlled comparison of two moxifloxacin containing treatment shortening regimens in pulmonary tuberculosis (REMox TB), registered at <http://clinicaltrials.gov/ct2/show/NCT00864383>, last accessed on 26 April 2010.
- [180] Rosenthal IM, Zhang M, Almeida D, Grosset JH, Nuermberger EL. Isoniazid or moxifloxacin in rifapentine-based regimens for experimental tuberculosis. *Am J Respir Crit Care Med* 2008; 178: 989 – 93.
- [181] Rifapentine plus moxifloxacin for treatment of pulmonary tuberculosis, registered at <http://clinicaltrials.gov/ct2/show/NCT00728507>; last accessed on 26 April 2010.
- [182] Agrawal D, Udwadia ZF, Rodriguez C, Mehta A. Increasing incidence of fluoroquinolone-resistant *Mycobacterium tuberculosis* in Mumbai, India. *Int J Tuberc Lung Dis* 2009; 13: 79 – 83.
- [183] Dheda K, Shean K, Zumla A, et al. Early Treatment Outcomes of Extensively Drug-Resistant Tuberculosis in South Africa are poor regardless of HIV status. *Lancet* [in press].
- [184] Cheng AF, Yew WW, Chan EW, Chin ML, Hui MM, Chan RC. Multiplex PCR amplicon conformation analysis for rapid detection of gyr A mutations in fluoroquinolone-resistant *Mycobacterium tuberculosis* clinical isolates. *Antimicrob Agents Chemother* 2004; 48: 596 – 601.
- [185] Andries K, Verhasselt P, Guillemont J, et al. A diarylquinoline drug active on the ATP synthase of *Mycobacterium tuberculosis*. *Science* 2005; 307: 223 – 7.
- [186] Lounis N, Gevers T, Van Den Berg J, Andries K. Impact of the interaction of R207910 with rifampin on the treatment of tuberculosis studied in the mouse model. *Antimicrob Agents Chemother* 2008; 52: 3568 – 72.

- [187] Ibrahim M, Andries K, Lounis N, et al. Synergistic activity of R207910 combined with pyrazinamide against murine tuberculosis. *Antimicrob Agents Chemother* 2007; 51: 1011 – 5.
- [188] Veziris N, Ibrahim M, Lounis N, et al. A once-weekly R207910-containing regimen exceeds activity of the standard daily regimen in murine tuberculosis. *Am J Respir Crit Care Med* 2009; 179: 75 – 9.
- [189] Lounis N, Veziris N, Chauffour A, Truffot-Pernot C, Andries K, Jarlier V. Combinations of R207910 with drugs used to treat multidrug-resistant tuberculosis have the potential to shorten treatment duration. *Antimicrob Agents Chemother* 2006; 50: 3543 – 7.
- [190] Rustomjee R, Diacon AH, Allen J, et al. Early bactericidal activity and pharmacokinetics of the diarylquinoline TMC-207 in treatment of pulmonary tuberculosis. *Antimicrob Agents Chemother* 2008; 52: 2831 – 5.
- [191] TMC207-TiDP13-C208: Anti-bacterial activity, safety, and tolerability of TMC207 in patients with multidrug-resistant *Mycobacterium tuberculosis* (MDR-TB), registered at <http://clinicaltrials.gov/ct2/show/NCT00449644>; last accessed on 26 April 2010.
- [192] Diacon AH, Pym A, Grobusch M, et al. The diarylquinoline TMC207 for multidrug-resistant tuberculosis. *N Engl J Med* 2009; 360: 2397 – 405.
- [193] Matsumoto M, Hashizume H, Tomishige T, et al. OPC-67683, a nitro-dihydro-imidazooxazole derivative with promising action against tuberculosis *in vitro* and in mice. *PLoS Med* 2006; 3: e466.
- [194] A placebo-controlled, phase 2 trial to evaluate OPC 67683 in patients with pulmonary sputum culture-positive, multidrug-resistant tuberculosis (TB), registered at <http://clinicaltrials.gov/ct2/show/NCT00685360>; last accessed at 26 April 2010.

- [195] Stover CK, Warrener P, VanDevanter DR, et al. A small-molecule nitroimidazopyran drug candidate for the treatment of tuberculosis. *Nature* 2000; 405: 962 – 6.
- [196] Tyagi S, Nuermberger E, Yoshimatsu T, et al. Bactericidal activity of the nitroimidazopyran PA-824 in a murine model of tuberculosis. *Antimicrob Agents Chemother* 2005; 49: 2289 – 93.
- [197] Tasneen R, Tyagi S, Williams K, Grosset J, Nuermberger E. Enhanced bactericidal activity of rifampin and/or pyrazinamide when combined with PA-824 in a murine model of tuberculosis. *Antimicrob Agents Chemother* 2008; 52: 3664 – 8.
- [198] Nuermberger E, Tyagi S, Tasneen R, et al. Powerful bactericidal and sterilizing activity of a regimen containing PA-824, moxifloxacin, and pyrazinamide in a murine model of tuberculosis. *Antimicrob Agents Chemother* 2008; 52: 1522 – 4.
- [199] Ginsberg AM, Laurenzi MW, Rouse DJ, Whitney KD, Spigelman MK. Safety, tolerability and pharmacokinetics of PA-824 in healthy subjects. *Antimicrob Agents Chemother* 2009; 53: 3720 – 5.
- [200] Diacon A, Dawson R, Venter A, et al. Update on PA-824. *Int J Tuberc Lung Dis* 2008; 12: S14.
- [201] Hu Y, Coates AR, Mitchison DA. Comparison of the sterilizing activities of the nitroimidazopyran PA-824 and moxifloxacin against persisting *Mycobacterium tuberculosis*. *Int J Tuberc Lung Dis* 2008; 12: 69 – 73.
- [202] Barry PJ, O'Connor TM. Novel agents in the management of *Mycobacterium tuberculosis* disease. *Curr Med Chem* 2007; 14: 2000 – 8.
- [203] Ma Z, Lienhardt C. Toward an optimized therapy for tuberculosis? *Drugs in clinical trials and in preclinical development. Clin Chest Med* 2009; 30: 755 – 68.

- [204] Amaral L, Boeree MJ, Gillespie SH, Udwadia ZF, van Soolingen D. Thioridazine cures extensively drug-resistant tuberculosis (XDR-TB) and the need for global trials is now. *Int J Antimicrob Agents* Feb [Epub ahead of print].
- [205] Iseman MD, Madsen L, Goble M, Pomerantz M. Surgical intervention in the treatment of pulmonary disease caused by drug-resistant *Mycobacterium tuberculosis*. *Am Rev Respir Dis* 1990; 141: 623 – 5.
- [206] Sihoe AD, Shiraishi Y, Yew WW. The current role of thoracic surgery in tuberculosis management. *Respirology* 2009; 14: 954 – 68.
- [207] Pomerantz M, Brown JM. Surgery in the treatment of multidrug-resistant tuberculosis. *Clin Chest Med* 1997; 18: 123 – 30.
- [208] Lalloo UG, Naidoo R, Ambaram A. Recent advances in the medical and surgical treatment of multidrug resistant tuberculosis. *Curr Opin Pulm Med* 2006; 12: 179 – 85.
- [209] Treasure RL, Seaworth BJ. Current role of surgery in *Mycobacterium tuberculosis*. *Ann Thorac Surg* 1995; 59: 1405 – 7.
- [210] van Leuven M, De Groot M, Shean KP, von Oppell UO, Willcox PA. Pulmonary resection as an adjunct in the treatment of multiple drug-resistant tuberculosis. *Ann Thorac Surg* 1997; 63: 1368 – 72.
- [211] Sung SW, Kang CH, Kim YT, Han SK, Shim YS, Kim JH. Surgery increased the chance of cure in multidrug resistant pulmonary tuberculosis. *Eur J Cardiothorac Surg* 1999; 16: 187 – 93.
- [212] Pomerantz BJ, Cleveland JC Jr, Olson HK, Pomerantz M. Pulmonary resection for multi-drug resistant tuberculosis. *J Thorac Cardiovasc Surg* 2001; 121: 448 – 53.
- [213] Chiang CY, Yu MC, Bai KJ, Suo J, Lin TP, Lee YC. Pulmonary resection in the treatment of patients with pulmonary multidrug-resistant tuberculosis in Taiwan. *Int J Tuberc Lung Dis* 2001; 5: 272 – 7.

- [214] Park SK, Lee CM, Heu JP, Song SD. A retrospective study for the outcome of pulmonary resection in 49 patients with multidrug-resistant tuberculosis. *Int J Tuberc Lung Dis* 2002; 6: 143 – 9.
- [215] Naidoo R, Reddi A. Lung resection for multidrug-resistant tuberculosis. *Asian Cardiovasc Thorac Ann* 2005; 13: 172 – 4.
- [216] Takeda S, Maeda H, Hayakawa M, Sawabata N, Maekura R. Current surgical intervention for pulmonary tuberculosis. *Ann Thorac Surg* 2005; 79: 959 – 63.
- [217] Kir A, Inci I, Torun T, Atasalihi A, Tahaoglu K. Adjunctive resectional surgery improves cure rates in multidrug-resistant tuberculosis. *J Thorac Cardiovasc Surg* 2006; 131: 693 – 6.
- [218] Somocurcio JG, Sotomayor A, Shin S, et al. Surgery for patients with drug-resistant tuberculosis: report of 121 cases receiving community-based treatment in Lima, Peru. *Thorax* 2007; 62: 416 – 21.
- [219] Kim HJ, Kang CH, Kim YT, et al. Prognostic factors for surgical resection in patients with multidrug-resistant tuberculosis. *Eur Respir J* 2006; 28: 576 – 80.
- [220] Mohsen T, Zeid AA, Haj-Yahia S. Lobectomy or pneumonectomy for multidrug-resistant pulmonary tuberculosis can be performed with acceptable morbidity and mortality: a seven-year review of a single institution's experience. *J Thorac Cardiovasc Surg* 2007; 134: 194 – 8.
- [221] Wang H, Lin H, Jiang G. Pulmonary resection in the treatment of multidrug-resistant tuberculosis: a retrospective study of 56 cases. *Ann Thorac Surg* 2008; 86: 1640 – 5.
- [222] Shiraishi Y, Katsuragi N, Kita H, Tominaga Y, Kariatsumari K, Onda T. Aggressive surgical treatment of multidrug-resistant tuberculosis. *J Thorac Cardiovasc Surg* 2009; 138: 1180 – 4.

- [223] Griffith DE. Treatment of multidrug-resistant tuberculosis. Should you try this at home? *Am J Respir Crit Care Med* 2004; 169: 1082 – 3.
- [224] Torun T, Tahaoglu K, Ozmen I, et al. The role of surgery and fluoroquinolones in the treatment of multidrug-resistant tuberculosis. *Int J Tuberc Lung Dis* 2007; 11: 979 – 85.
- [225] Park SK, Kim JH, Kang H, Cho JS, Smego RA Jr. Pulmonary resection combined with rifampin-based drug therapy for patients with multidrug-resistant and extensively drug-resistant tuberculosis. *Int J Infect Dis* 2009; 13: 170 – 5.
- [226] Kwon YS, Kim YH, Suh GY, et al. Treatment outcomes for HIV-uninfected patients with multidrug-resistant and extensively drug-resistant tuberculosis. *Clin Infect Dis* 2008; 47: 496 – 502.
- [227] Dravniece G, Cain KP, Holtz TH, Riekstina V, Leimane V, Zaleskis R. Adjunctive resectional lung surgery for extensively drug-resistant tuberculosis. *Eur Respir J* 2009; 34: 180 – 3.
- [228] Shiraishi Y, Katsuragi N, Kita H, Toishi M, Onda T. Experience with pulmonary resection for extensively drug-resistant tuberculosis. *Interact Cardiovasc Thorac Surg* 2008; 7: 1075 – 8.
- [229] Kim DH, Kim HJ, Park SK, et al. Treatment outcomes and long-term survival in patients with extensively drug-resistant tuberculosis. *Am J Respir Crit Care Med* 2008; 178: 1075 – 82.
- [230] Naidoo R. Surgery for pulmonary tuberculosis. *Curr Opin Pulm Med* 2008; 14: 254 – 9.
- [231] Dewan RK, Singh S, Kumar A, Meena BK. Thoracoplasty: an obsolete procedure? *Indian J Chest Dis Allied Sci* 1999; 41: 83 – 8.

- [232] Jouveshomme S, Dautzenberg B, Bakdach H, Derenne JP. Preliminary results of collapse therapy with plombage for pulmonary disease caused by multidrug-resistant mycobacteria. *Am J Respir Crit Care Med* 1998; 157: 1609 – 15.
- [233] Motus IY, Skorniakov SN, Sokolov VA, et al. Reviving an old idea: can artificial pneumothorax play a role in the modern management of tuberculosis? *Int J Tuberc Lung Dis* 2006; 10: 571 – 7.
- [234] Jouanguy E, Doffinger R, Dupuis S, Pallier A, Altare F, Casanova JL. IL-12 and IFN-gamma in host defense against mycobacteria and salmonella in mice and men. *Curr Opin Immunol* 1999; 11: 346 – 51.
- [235] Sugawara I, Yamada H, Kaneko H, Mizuno S, Takeda K, Akira S. Role of interleukin-18 (IL-18) in mycobacterial infection in IL-18-gene-disrupted mice. *Infect Immun* 1999; 67: 2585 – 9.
- [236] Fortes A, Pereira K, Antas PR, et al. Detection of *in vitro* interferon-gamma and serum tumour necrosis factor-alpha in multidrug-resistant tuberculosis patients. *Clin Exp Immunol* 2005; 141: 541 – 8.
- [237] Rook GA. Th2 cytokines in susceptibility to tuberculosis. *Curr Mol Med* 2007; 7: 327 – 37.
- [238] Churchyard GJ, Kaplan G, Fallows D, Wallis RS, Onyebujoh P, Rook GA. Advances in immunotherapy for tuberculosis treatment. *Clin Chest Med* 2009; 30: 769-82.
- [239] Mayosi BM, Volmink JA, Commerford PJ. Interventions for treating tuberculous pericarditis. *Cochrane Database Syst Rev* 2000; 2: CD000526.
- [240] Prasad K, Singh MB. Corticosteroids for managing tuberculous meningitis. *Cochrane Database Syst Rev* 2008; 1: CD002244.
- [241] Matchaba PT, Volmink J. Steroids for treating tuberculous pleurisy. *Cochrane Database Syst Rev* 2000; 2: CD001876.

- [242] Thwaites GE, Nguyen DB, Nguyen HD, et al. Dexamethasone for the treatment of tuberculous meningitis in adolescents and adults. *N Engl J Med* 2004; 351: 1741 – 51.
- [243] Elliott AM, Luzze H, Quigley MA, et al. A randomized double-blind, placebo-controlled trial of the use of prednisolone as an adjunct to treatment in HIV-associated pleural tuberculosis. *J Infect Dis* 2004; 190: 869 – 78.
- [244] Mayanja-Kizza H, Jones-Lopez E, Okwera A, et al. Immunoadjuvant prednisolone therapy for HIV-associated tuberculosis: a phase 2 clinical trial in Uganda. *J Infect Dis* 2005; 191: 856 – 65.
- [245] Condos R, Schluger NW. Cytokine-based approaches to the treatment of multidrug-resistant tuberculosis. *Bio Drugs* 1999; 11: 165 – 73.
- [246] Condos R, Rom WN, Schluger NW. Treatment of multidrug-resistant pulmonary tuberculosis with interferon-gamma via aerosol. *Lancet* 1997; 349: 1513 – 5.
- [247] Johnson BJ, Bekker LG, Rickman R, et al. rhuIL-2 adjunctive therapy in multidrug-resistant tuberculosis: a comparison of two treatment regimens and placebo. *Tuber Lung Dis* 1997; 78: 195 – 203.
- [248] Palmero D, Eiguchi K, Rendo P, Castro Zorrilla L, Abbate E, Gonzalez Montaner LJ. Phase II trial of recombinant interferon-alpha2b in patients with advanced intractable multidrug-resistant pulmonary tuberculosis: long-term follow-up. *Int J Tuberc Lung Dis* 1999; 3: 214 – 8.
- [249] Giosue S, Casarini M, Ameglio F, et al. Aerosolized interferon-alpha treatment in patients with multidrug-resistant pulmonary tuberculosis. *Eur Cytokine Netw* 2000; 11: 99 – 104.
- [250] Grahmann PR, Braun RK. A new protocol for multiple inhalation of IFN-gamma successfully treats MDR-TB: a case study. *Int J Tuberc Lung Dis* 2008; 12: 636 – 44.

- [251] Park SK, Cho S, Lee IH, et al. Subcutaneously administered interferon-gamma for the treatment of multidrug-resistant pulmonary tuberculosis. *Int J Infect Dis* 2007; 11: 434 – 40.
- [252] Stanford JL, Stanford CA, Grange JM, Lan NN, Etemadi A. Does immunotherapy with heat-killed *Mycobacterium vaccae* offer hope for the treatment of multidrug-resistant pulmonary tuberculosis? *Respir Med* 2001; 95: 444 – 7.
- [253] Dawson R, Condos R, Tse D, et al. Immunomodulation with recombinant interferon-gamma1b in pulmonary tuberculosis. *PLoS One*. 2009;4:e6984.
- [254] Ralph AP, Kelly PM, Anstey NM. L-arginine and vitamin D: novel adjunctive immunotherapies. *Trends Microbiol* 2008; 16: 336 – 44.
- [255] Wejse C, Gomes VF, Rabna P, et al. Vitamin D as supplementary treatment for tuberculosis: a double-blind, randomized placebo-controlled trial. *Am J Respir Crit Care Med* 2009; 179: 843 – 50.
- [256] Martins M, Viveiros M, Couto I, Amaral L. Targeting human macrophages for enhanced killing of intracellular XDR-TB and MDR-TB. *Int J Tuberc Lung Dis* 2009; 13: 569 – 73.
- [257] Migliori GB, Loddenkemper R, Blasi F, Raviglione MC. 125 years after Robert Koch's discovery of the tubercle bacillus: the new XDR-TB threat. Is "science" enough to tackle the epidemic. *Eur Respir J* 2007; 29: 423 – 7.